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DNA isolates encoding the human C-C chemokine receptor C-C CKR-1 and methods of obtaining such DNA are provided, together with expression systems for recombinant production of C-C CKR-1 useful in therapeutic or diagnostic compositions. Additionally, a method for identifying new chemokine receptors is provided.

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C-C CKR-1, C-C CHEMOKINE RECEPTOR

This invention relates to the field of cytokine receptors, their antibodies, and their use as diagnostic and therapeutic agents.

BACKGROUND OF THE INVENTION

Cytokines are biological molecules which affect inflammatory and immune-related effector cells. The inflammatory cytokines, or "chemokines," have a variety of biological properties including selective leukocyte chemotaxis and activation. These chemokines form a superfamily, denoted in the literature alternatively as the PF4 superfamily or the intercrines, that has been divided into two classes based on whether the first two conserved cysteine residues are separated by an intervening amino acid (C-X-C, or α), or whether they are adjacent (C-C, or β). The C-X-C class members include, for example, interleukin-8 (IL-8), melanocyte growth stimulating factor (MGSA), and platelet factor 4 (PF4), while the C-C class includes RANTES (Regulated on Activation, Normal T Expressed and Secreted) and monocyte chemotactic peptide-1 (MCP-1). The C-X-C class exerts proinflammatory activity mainly through their action on neutrophils, whereas the C-C class appears to be monocyte chemoattractants.

Much attention has been focused on receptor/ligand interactions in this superfamily, with more data being available on C-X-C than C-C chemokine binding. It has become clear that chemokine receptor/ligand interactions on target inflammatory cells seem to be strictly regulated. For example, no cross competition for binding sites has been observed on either monocytes or neutrophils between members of the C-X-C or C-C branches (Leonard, E. J. et al. Immunol. Today 11:97-101, 1990; Samanta, A. K. et al. J. Exp. Med. 169:1185-1189, 1989; Yoshimura, T. et al. J. Immunol. 145:292-7, 1990), consistent with the differential chemoattractant effects on these two cell types. Direct binding data for the C-C chemokines is surprisingly sparse.

Human MCP-1 has been reported to bind to monocytes with an affinity of about 2 nM, with no sites detectable on neutrophils (Valente, A. J. et al. Biochem. Biophys. Res. Commun. 176:309-14, 1991; Yoshimura, T. et al. J. Immunol. 145:292-7, 1990). A single report shows human Act-2, a human MIP-1 β (HuMIP-1 β) variant, binding to between 7,000 and 45,000 sites on PBMC with an affinity of between 7.8 and 12 nM (Napolitano, M. et al. J. Exp. Med. 172:285-289, 1990). Kwon and colleagues have characterized the binding of murine MIP-1 α on a mouse T cell and a macrophage cell line, finding a K_d of 1.5 and 0.9 nM, respectively (Oh, K. O. et al. J. Immunol. 147:2978-83, 1991).

Most of the molecular details regarding leukocyte motility remain to be elucidated. Recently, however, the receptors for the anaphylatoxin C5a (Gerard, N. P. et al. Nature 349:614-7, 1991), the bacterial formylated tripeptide fMLP (Boulay, F. et al. Biochemistry 29:11123-11133, 1990), and the C-X-C chemokine IL-8 (Holmes, W. E. et al. Science 253:1278-80, 1991; Murphy, P. M. et al. Science 253:1280-3, 1991) have been cloned using molecular techniques. All of these receptors display amino acid sequences which are predicted to conform to an architecture containing seven-transmembrane-spanning segments connected by a series of intra- and extracellular loops. The primary sequences of these receptors revealed domains which were conserved in receptors associated with cell motility, but not in other seven-transmembrane-spanning receptors.

Accordingly, it is an object of the invention to provide isolated C-C chemokine receptor (C-C CKR-1) for use as a therapeutic or diagnostic reagent.

It is another object of the invention to make variants of C-C CKR-1 for use as antagonists or agonists.

It is another object of the invention to generate antibodies against C-C CKR-1 for use as diagnostic and therapeutic agents.

It is another object of the invention to provide a method for identifying new chemokine receptors.

SUMMARY OF THE INVENTION

One aspect of the invention is the isolation of the novel chemokine receptor, C-C CKR-1.

In another aspect, the invention provides a
5 composition comprising C-C CKR-1 that is free of contaminating polypeptides of the animal species from which the C-C CKR-1 is derived.

In another aspect of the invention, C-C CKR-1, or fragments thereof (which also may be synthesized by
10 chemical methods), is fused (by recombinant expression or in vitro covalent methods) to an immunogenic polypeptide and this fusion polypeptide, in turn, is used to immunize an animal to raise antibodies against a C-C CKR-1 epitope. Anti-C-C CKR-1 antibodies are recovered from the serum of
15 immunized animals. Alternatively, monoclonal antibodies are prepared from cells of the immunized animal in conventional fashion.

Another aspect of the invention is the use of anti-C-C CKR-1 antibodies in the diagnosis of (in vitro or in vivo)
20 or (when immobilized on an insoluble matrix) the purification of chemokine receptors which bind thereto.

Another aspect of the invention is the derivatization of C-C CKR-1 in vitro to prepare immobilized C-C CKR-1 and labeled C-C CKR-1, particularly for purposes of diagnosis
25 of C-C CKR-1 or its antibodies, or for affinity purification of C-C CKR-1 antibodies themselves.

Another aspect of the invention is the formulation of C-C CKR-1, its derivatives, or its antibodies into physiologically acceptable vehicles, especially for
30 therapeutic use. Such vehicles include sustained-release formulations of C-C CKR-1.

In still other aspects, the invention provides an isolated nucleic acid molecule encoding C-C CKR-1, labeled or unlabeled, and a nucleic acid sequence that is
35 complementary to, or hybridizes under defined conditions to a nucleic acid sequence encoding C-C CKR-1.

In addition, the invention provides a replicable vector comprising the nucleic acid molecule encoding C-C CKR-1 operably linked to control sequences recognized by a

host transformed by the vector; host cells transformed with the vector; and a method of using a nucleic acid molecule encoding C-C CKR-1 to effect the production of C-C CKR-1, comprising expressing the nucleic acid molecule in a culture of the transformed host cells and recovering C-C CKR-1 from the host cell culture. The nucleic acid sequence is also useful in hybridization assays for C-C CKR-1 nucleic acid.

Another aspect of the invention is substitutional, deletional, or insertional variants of C-C CKR-1 amino acids and/or glycosyl residues, including variants having non-native glycosylation. These variants are prepared by in vitro or recombinant methods. Sequence variants are optionally screened for immuno-cross-reactivity with C-C CKR-1 and for C-C CKR-1 antagonist or agonist activity.

Another aspect of the invention is a method for identifying new C-C chemokine receptors.

Another aspect of the invention is a method for determining the biological activity of a C-C chemokine variant on C-C CKR-1, by transforming a host cell with DNA encoding C-C CKR-1, culturing the host cell to express the receptor on its surface, harvesting the cells, contacting the cells with a C-C chemokine variant, and determining the biological activity of the variant on the receptor.

In further embodiments, the invention provides transgenic animals comprising C-C CKR-1 from another species, animals in which C-C CKR-1 is expressed in a tissue in which it is not ordinarily found, or animals in which C-C CKR-1 is inactivated, by, for example, gene disruption.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 (SEQ ID NOS:1-7) depicts the predicted amino acid sequence of C-C CKR-1, and alignment of the predicted amino acid sequence with HUMSTSR (Genbank accession #M99293), IL8rA (Holmes, W. E. et al. Science 253:1278-80, 1991), IL8rB (Murphy, P. M. et al. Science 253:1280-3, 1991), C5a receptor (Gerard, N. P. et al. Nature 349:614-7, 1991), fMLP receptor (Boulay, F. et al. Biochemistry

29:11123-11133, 1990) and the open reading frame of cytomegalovirus, US28. The seven putative transmembrane spanning domains are overlined. Glycosylation sites are indicated with black dots above the sequence. The two cysteine residues implicated in disulfide bonding are indicated with asterisks. The consensus site for protein kinase C phosphorylation is indicated as "+". Conserved amino acids appearing more than two times in the alignment are boxed.

10 Figure 2 depicts Northern blot analysis of RNA from hematopoietic cells probed with C-C CKR-1. 5 µg of poly A⁺ or 20 µg of total RNA from the cell lines indicated was size fractionated on 1% formaldehyde-agarose, transferred to nitrocellulose and hybridized with radiolabeled C-C CKR-1 cDNA. The filter was washed with 0.5X SSC, 0.1% SDS at 55°C and the autoradiograph was developed after 4-6 hours exposure at -70°C with intensifying screens. RNA molecular weight markers were also run on the gel and are indicated on the side.

20 Figures 3A and 3B depict Southern blot analysis of human genomic DNA probed with C-C CKR-1. In Figure 3A, 10 µg of genomic DNA was digested with the restriction enzyme indicated, run on a 0.6% agarose gel, blotted onto Genescreen® and hybridized with radiolabeled C-C CKR-1 cDNA. The filter was washed with 0.5X SSC, 1% SDS at 55°C and the autoradiograph was developed after overnight exposure at -70°C with intensifying screens. DNA molecular weight markers were also run on the gel and are indicated on the side. In Figure 3B, the same blot was washed more stringently with 0.2X SSC, 0.1% SDS at 60°C and autoradiography performed as indicated before.

 Figures 4A and 4B are graphs depicting intracellular Ca⁺⁺ concentrations of 293 cells transfected with C-C CKR-1 cDNA and challenged with human MIP-1α (HuMIP-1α) and RANTES. In Figure 4A, 50% confluent cells were transfected with 10-20 µg of plasmid DNA by the calcium-phosphate precipitation method. After transient expression for 12-24 hours, cells were harvested, loaded with the calcium probe INDO-1 AM and

assayed by spectrofluorometric methods at 37°C with continuous stirring. Various concentrations of HuMIP-1 α , as indicated, were added after 12 seconds. The intracellular concentrations of Ca⁺⁺ was determined as described (Naccache, P. H. et al. J. Immunol. 142:2438-44, 1989). In Figure 4B, details were as for Figure 4A, except that various concentration of RANTES, as indicated, were used.

Figures 5A-5D are graphs depicting desensitization in response to the challenge of the same or different ligands by 293 cells transiently expressing C-C CKR-1. Details are as described in Figure 4. The transfected cells were first challenged at 12 seconds with 100 nM of HuMIP-1 α or 250 nM of RANTES, and then at 70 seconds with the same concentration of ligands in the order indicated.

Figures 6A and 6B are graphs depicting the binding of ¹²⁵I-HuMIP-1 α and ¹²⁵I-RANTES on 293 cells transfected with C-C CKR-1 cDNA. In Figure 6A, Human embryonic kidney cells (293 cells) were transfected with 10-20 μ g plasmid DNA as described in Figure 4. Transfected cells were incubated for 2 hours at 4°C with ¹²⁵I-HuMIP-1 α in the presence of increasing concentrations of unlabeled HuMIP-1 α . The inset shows Scatchard analysis of the binding data and revealed a K_d of 5.1 \pm 0.3nM for ¹²⁵I-MIP-1 α to C-C CKR-1. Figure 6B depicts displacement of ¹²⁵I-RANTES with unlabeled HuMIP-1 α on 293 cells transfected with the C-C CKR-1 cDNA. Scatchard analysis of the binding data revealed a K_d of 7.6 \pm 1.5nM for the displacement of ¹²⁵I-RANTES to the C-C CKR-1.

Figure 7 is a graph depicting displacement of ¹²⁵I-HuMIP-1 α binding to 293 cells transfected with C-C CKR-1 cDNA. Cells were transfected as outlined in Figure 4 and incubated for 2 hours at 4°C with ¹²⁵I-HuMIP-1 α in the presence of increasing concentrations of the cross competing ligands, HuMIP-1 α , murine MIP-1 α , HuMIP-1 β , MCP-1 and IL-8. The K_d and the number of sites, shown in the bottom left corner, were determined by Scatchard analysis of the binding data.

Figure 8 is a graph depicting the intracellular Ca^{++} concentration of 293 cells transiently expressing C-C CKR-1 and challenged with HuMIP-1 α , RANTES, HuMIP-1 β and MCP-1. Details are as described in Figure 4.

5 Figure 9 (SEQ ID NO:8) is the nucleotide sequence of C-C CKR-1 and its 3' noncoding region.

Figure 10 is a graph depicting the binding of radiolabeled HuMIP-1 α to 293 cells transfected with the coding region of the open reading frame US28 in the
10 cytomegalovirus (CMV) genome. 293 cells were transfected with an expression construct containing the coding sequence of US28 in the sense or antisense orientation. After 12 hours, the cells were harvested and incubated with 0.9 nM of ^{125}I -HuMIP-1 α in combination with 1 μM of either
15 unlabeled HuMIP-1 α , murine MIP-1 β , MCP-1, RANTES, or IL-8. The amount of displaceable ^{125}I -HuMIP-1 α was determined by subtracting the amount of ^{125}I -HuMIP-1 α bound in the absence of any cold ligand from the amount bound in the presence of cold ligand. Background refers to counts
20 obtained from cells transfected with the antisense orientation of US28.

DETAILED DESCRIPTION OF THE INVENTION

A. Definitions

25

In general, the following words or phrases have the indicated definition when used in the description, examples, and claims.

"C-C CKR-1" is the chemokine receptor described infra
30 together with its amino acid sequence or cellular analogs, alleles, predetermined amino acid sequence mutations, glycosylation variants, and covalent modifications. Embodiments of C-C CKR-1 exclude known chemokine receptors, in particular those which are set forth in the Background
35 section above, and chemokine receptors statutorily obvious from those such chemokine receptors.

"Orphan receptor" is defined as the predicted polypeptide encoded by nucleic acid which hybridizes under low stringency conditions to probes designed from known

cytokine receptor nucleic acid sequences or other known sequences likely to have structural similarity to cytokine receptors, or detectable by PCR primers so designed, wherein the predicted polypeptide is not previously known
5 in the art.

"C-C CKR-1 qualitative biological activity" is defined as immunological cross-reactivity with at least one epitope of purified C-C CKR-1.

"Immunologically cross-reactive" is intended to mean
10 that the candidate polypeptide is capable of competitively inhibiting the binding of native C-C CKR-1 to polyclonal antibodies or antisera raised against native C-C CKR-1, respectively.

"Isolated C-C CKR-1 nucleic acid or polypeptide" is a
15 C-C CKR-1 nucleic acid or polypeptide that is identified and separated from at least one contaminant (nucleic acid or polypeptide respectively) with which it is ordinarily associated in nature, such as from the human source of C-C CKR-1 nucleic acid or polypeptide. In preferred
20 embodiments, C-C CKR-1 will be isolated to pharmaceutically acceptable levels of purity with respect to proteins of its species of origin. In preferred embodiments, C-C CKR-1 protein will be purified (1) to greater than 95% by weight of protein, and most preferably more than 99% by weight,
25 (2) to a degree sufficient to obtain at least 15 residues of N-terminal or internal amino acid sequence by an amino acid sequenator commercially available on the filing date hereof, or (3) to homogeneity by conventional nonreducing SDS polyacrylamide gel electrophoresis (SDS-PAGE) using
30 Coomassie blue or, preferably, silver stain. Isolated C-C CKR-1 includes C-C CKR-1 in situ within recombinant cells which do not ordinarily express the C-C CKR-1 in question, since, in this instance, at least one component of C-C CKR-1 natural environment will not be present. Isolated C-C
35 CKR-1 includes C-C CKR-1 in a recombinant cell culture of another species than the species of origin of the C-C CKR-1 since the C-C CKR-1 in such circumstances will be devoid of source polypeptides. Ordinarily, however, isolated C-C CKR-1 will be prepared by at least one purification step.

Isolated C-C CKR-1 nucleic acid includes a nucleic acid that is identified and separated from at least one containment nucleic acid with which it is ordinarily associated in the natural source of the C-C CKR-1 nucleic acid. Isolated C-C CKR-1 nucleic acid thus is present in other than in the form or setting in which it is found in nature. However, isolated C-C CKR-1-encoding nucleic acid includes C-C CKR-1 nucleic acid in ordinarily C-C CKR-1-expressing cells where the nucleic acid is in a chromosomal location different from that of natural cells or is otherwise flanked by a different DNA sequence than that found in nature.

The nucleic acid or polypeptide may be labeled for diagnostic and probe purposes, using a label as described and defined further below in the discussion of diagnostic assays.

"C-C CKR-1 nucleic acid" is defined as RNA or DNA (a) containing at least 25 bases of the genomic or cDNA sequence that encodes C-C CKR-1, (b) is complementary to the genomic or cDNA sequence that encodes C-C CKR-1, (c) which hybridizes to such nucleic acid and remains stably bound to it under stringent conditions, or (d) encodes a polypeptide sharing at least 50% sequence identity over the entire length of the polypeptide, preferably at least 60%, and more preferably at least 70%, with the amino acid sequence of C-C CKR-1, and which polypeptide has the ability to bind at least one C-C chemokine. Preferably the hybridizing RNA or DNA contains at least 25 bases, more preferably 40, and more preferably 60 bases which are identical to the sequences encoding the C-C CKR-1 described infra. Optimally, C-C CKR-1 nucleic acid consists essentially only of sequence encoding C-C CKR-1 or the complement of such sequences.

"Stringency" conditions for hybridization are defined by washing conditions after the hybridization reaction. Typically, hybridization conditions are defined as employing overnight incubation at 42°C, in a solution comprising 20% formamide, 5X SSC (150 mM NaCl, 15 mM trisodium citrate), 50 mM sodium phosphate (pH 7.6), 5X

Denhardt's solution, 10% dextran sulfate, and 20 µg/ml denatured, sheared salmon sperm DNA. "High stringency" conditions for washing are defined as typically employing 0.2X SSC, 0.1% SDS at 55°C, while "low stringency"

5 conditions for washing are defined as typically employing 0.5X SSC, 1% SDS at 42°C. These conditions are well known in the art. See, for example, Current Protocols in Molecular Biology, eds. Ausubel, et al., Greene Publishing Associates, NY, 1989.

10 The term "control sequences" refers to DNA sequences necessary for the expression of an operably linked coding sequence in a particular host organism. The control sequences that are suitable for prokaryotes, for example, include a promoter, optionally an operator sequence, and a
15 ribosome binding site. Eukaryotic cells are known to utilize promoters, polyadenylation signals, and enhancers.

Nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For example, DNA for a presequence or secretory
20 leader is operably linked to DNA for a polypeptide if it is expressed as a preprotein that participates in the secretion of the polypeptide; a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence; or a ribosome binding site
25 is operably linked to a coding sequence if it is positioned so as to facilitate translation. Generally, "operably linked" means that the DNA sequences being linked are contiguous and, in the case of a secretory leader, contiguous and in reading phase. However enhancers do not
30 have to be contiguous. Linking is accomplished by ligation at convenient restriction sites. If such sites do not exist, then synthetic oligonucleotide adapters or linkers are used in accord with conventional practice.

The starting plasmids used to practice this invention
35 are commercially available, are publicly available on an unrestricted basis, or can be constructed from such available plasmids in accord with published procedures. In addition, other equivalent plasmids are known in the art and will be apparent to the ordinary artisan. Methods for

restriction enzyme digestion, recovery or isolation of DNA, hybridization analysis, and ligation are conventional and by this time well known to the ordinary artisan.

Similarly, the cell lines used to practice this invention
5 are commercially available or are publicly available on an unrestricted basis.

Another method for obtaining the gene of interest is to chemically synthesize it using one of the methods described in Engels et al. (Agnew, Chem. Int. Ed. Engl.
10 28:716-734, 1989). These methods include triester, phosphite, phosphoramidite and H-phosphonate methods, typically proceeding by oligonucleotide synthesis on solid supports.

"Recovery" or "isolation" of a given fragment of DNA
15 from a restriction digest means separation of the digest on polyacrylamide or agarose gel by electrophoresis, identification of the fragment of interest by comparison of its mobility versus that of marker DNA fragments of known molecular weight, removal of the gel section containing the
20 desired fragment, and separation of the gel from DNA. This procedure is known generally. For example, see Sambrook, et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY 1989).

Amino acids are referred to by their standard three
25 letter IUPAC abbreviations.

B. General Methods for Practicing the Invention

1. Preparation of Native C-C CKR-1 Nucleic Acid

The use of the singular article "the" with respect to
30 C-C CKR-1 is not intended to suggest that only one DNA sequence encodes C-C CKR-1. In fact, it is expected that alleles, processing intermediates, and predetermined sequence variants described infra will vary in sequence from the DNA encoding native C-C CKR-1. Further, C-C CKR-1
35 may fall within a subfamily of chemokine receptors having a high degree of sequence homology but which vary sufficiently as to not constitute alleles. All of these sequences fall within the ambit of C-C CKR-1 nucleic acid.

DNA sequences encoding C-C CKR-1 may be either genomic

or cDNA. Any representative genomic library may be screened with the probes described below. Methods for genomic DNA preparation and the construction of cDNA libraries are well known in the art. See, for example,
5 Sambrook et al., supra.

2. Amino Acid Sequence Variants of C-C CKR-1

Amino acid sequence variants of C-C CKR-1 are prepared by introducing appropriate nucleotide changes into C-C CKR-
10 1 DNA, or by in vitro synthesis of the desired C-C CKR-1 polypeptide. Such variants include, for example, deletions from, or insertions or substitutions of, residues within the amino acid sequence of native C-C CKR-1. Any combination of deletion, insertion, and substitution can be
15 made to arrive at the final construct, provided that the final construct possesses the desired characteristics.

The amino acid changes also may alter post-translational processing of C-C CKR-1, such as changing the number or position of glycosylation sites or by altering
20 its membrane anchoring characteristics.

In designing amino acid sequence variants of C-C CKR-1, the location of the mutation site and the nature of the mutation will depend on C-C CKR-1 characteristic(s) to be modified. The sites for mutation can be modified
25 individually or in series, e.g., by (1) substituting first with conservative amino acid choices and then with more radical selections depending upon the results achieved, (2) deleting the target residue, or (3) inserting residues of the same or a different class adjacent to the located site,
30 or combinations of options 1-3.

A useful method for identification of certain residues or regions of C-C CKR-1 polypeptide that are preferred locations for mutagenesis is called "alanine scanning mutagenesis" as described by Cunningham and Wells (Science
35 244:1081-1085, 1989). Here, a residue or group of target residues are identified (e.g., charged residues such as arg, asp, his, lys, and glu) and replaced by a neutral or negatively charged amino acid (most preferably alanine or polyalanine) to affect the interaction of the amino acids

with the surrounding aqueous environment in or outside the cell. Those domains demonstrating functional sensitivity to the substitutions then are refined by introducing further or other variants at or for the sites of substitution. Thus, while the site for introducing an amino acid sequence variation is predetermined, the nature of the mutation per se need not be predetermined. For example, to optimize the performance of a mutation at a given site, alanine scanning or random mutagenesis may be conducted at the target codon or region and the expressed C-C CKR-1 variants are screened for the optimal combination of desired activity.

C-C CKR-1 variants will exhibit at least a biological activity of the parental sequence, for example, chemokine binding or antigenic activity. Preferably, the antigenically active C-C CKR-1 is a polypeptide that binds to an antibody raised against the polypeptide in its native conformation, "native conformation" generally meaning the polypeptide as found in nature which has not been denatured by chaotropic agents, heat or other treatment that substantially modifies the three dimensional structure of the polypeptide (this can be determined, for example, by migration on nonreducing, nondenaturing sizing gels). Antibody used in determination of antigenic activity is rabbit polyclonal antibody raised by formulating the native non-rabbit polypeptide in Freund's complete adjuvant, subcutaneously injecting the formulation, and boosting the immune response by intraperitoneal injection of the formulation until the titer of anti-polypeptide antibody plateaus.

Amino acid sequence deletions generally range from about 1 to 30 residues, more preferably about 1 to 10 residues, and typically are contiguous. Preferably, deletions are made in regions of the protein that are the least conserved when C-C CKR-1 amino acid sequence is compared with other chemokine receptors. Such deletions will be more likely to modify the biological activity of the polypeptides more significantly than deletions made elsewhere. The number of consecutive deletions will be

selected so as to preserve the tertiary structure of C-C CKR-1 in the affected domain, e.g., beta pleated sheet or alpha helix.

Amino acid sequence insertions include amino- and/or
5 carboxyl-terminal fusions ranging in length from one
residue to polypeptides containing a hundred or more
residues, as well as intrasequence insertions of single or
multiple amino acid residues. Intrasequence insertions
(i.e., insertions within C-C CKR-1 sequence) may range
10 generally from about 1 to 10 residues, more preferably 1 to
5, most preferably 1 to 3.

Insertional variants of C-C CKR-1 or its extracellular
segments include the fusion to the N- or C-terminus of C-C
CKR-1 of immunogenic polypeptides, e.g., bacterial
15 polypeptides such as β -lactamase or an enzyme encoded by
the E. coli trp locus, or yeast protein, and C-terminal
fusions with proteins having a long half-life such as in
place of V_H or V_C domains of immunoglobulins comprising
constant regions, albumin, or ferritin (for example, as
20 described in WO 89/02922, published 6 April 1989).

Another group of variants are amino acid substitution
variants. These variants have at least one amino acid
residue in C-C CKR-1 molecule removed and a different
residue inserted in its place. The sites of greatest
25 interest for substitutional mutagenesis include sites
identified as the active site(s) of C-C CKR-1, and sites
where the amino acids found in C-C CKR-1 from various
species are substantially different in terms of side-chain
bulk, charge, and/or hydrophobicity.

30 Other sites of interest are those in which particular
residues of C-C CKR-1 are conserved when compared with
other chemokine receptors. These positions may be
important for the biological activity of C-C CKR-1. These
sites, especially those falling within a sequence of at
35 least three other identically conserved sites, are
substituted in a relatively conservative manner. Such
conservative substitutions are shown in Table I under the
heading of preferred substitutions. If such substitutions
result in a change in biological activity, then more

substantial changes, denominated exemplary substitutions in Table I, or as further described below in reference to amino acid classes, are introduced and the products screened.

5

Table I

	<u>Original Residue</u>	<u>Exemplary Substitutions</u>	<u>Preferred Substitutions</u>
10	Ala (A)	ser	ser
	Arg (R)	lys; gln; asn; ala	lys
	Asn (N)	gln; his; lys; arg; ala; asp	asp
15	Asp (D)	glu; asn; ala	asn
	Cys (C)	ser; ala; val	ala
	Gln (Q)	asn; glu; ala	asn; glu
	Glu (E)	asp; gln; ala	gln
	Gly (G)	ala; asn	ala
20	His (H)	asn; gln; lys; arg; ala	asn
	Ile (I)	leu; val; met; ala; phe	val
	Leu (L)	ile; val; met; ala; phe	met
	Lys (K)	arg; gln; asn; met; ala	arg
25	Met (M)	leu; phe; ile; ala	leu
	Phe (F)	leu; val; ile; ala; tyr	leu
	Pro (P)	ala	ala
	Ser (S)	thr; ala	ala
	Thr (T)	ser; val; ala	ser
30	Trp (W)	tyr; phe; ala	tyr
	Tyr (Y)	trp; phe; thr; ala; gln	phe
	Val (V)	ile; leu; met; phe; ala; thr	ala

35 Substantial modifications in function or immunological identity of C-C CKR-1 are accomplished by selecting substitutions that differ significantly in their effect on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a sheet or
40 helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the

side chain. Naturally occurring residues are divided into groups based on common side chain properties:

- (1) hydrophobic: norleucine, met, ala, val, leu, ile;
- (2) neutral hydrophilic: cys, ser, thr;
- 5 (3) acidic: asp, glu;
- (4) basic: asn, gln, his, lys, arg;
- (5) residues that influence chain orientation: gly, pro;
- and
- (6) aromatic: trp, tyr, phe.

10 Non-conservative substitutions will entail exchanging a member of one of these classes for another. Such substituted residues may be introduced into regions of C-C CKR-1 that are homologous with other chemokine receptors, or, more preferably, into the non-homologous regions of the
15 molecule.

Any cysteine residues not involved in maintaining the proper conformation of C-C CKR-1 may be substituted, generally with serine, to improve the oxidative stability of the molecule and prevent aberrant cross-linking.

20 DNA encoding amino acid sequence variants of C-C CKR-1 is prepared by a variety of methods known in the art. These methods include, but are not limited to, isolation from a natural source (in the case of naturally occurring amino acid sequence variants) or preparation by
25 oligonucleotide-mediated (or site-directed) mutagenesis, PCR mutagenesis, and cassette mutagenesis of an earlier prepared variant or a non-variant version of C-C CKR-1. These techniques may utilize C-C CKR-1 nucleic acid (DNA or RNA), or nucleic acid complementary to C-C CKR-1 nucleic
30 acid. Oligonucleotide-mediated mutagenesis is a preferred method for preparing substitution, deletion, and insertion variants of C-C CKR-1 DNA. This technique is well known in the art (see, for example, as described by Adelman et al., DNA 2:183, 1983). PCR mutagenesis is also suitable for
35 making amino acid variants of C-C CKR-1 (see Erlich, supra, pp. 61-70). Another method for preparing variants, cassette mutagenesis, is based on the technique described by Wells et al. (Gene 34:315-323, 1985).

3. Insertion of DNA into a Cloning Vehicle

The cDNA or genomic DNA encoding native or variant C-C CKR-1 is inserted into a replicable vector for further cloning (amplification of the DNA) or for expression. Many
5 vectors are available, and selection of the appropriate vector will depend on (1) whether it is to be used for DNA amplification or for DNA expression, (2) the size of the DNA to be inserted into the vector, and (3) the host cell to be transformed with the vector. Each vector contains
10 various components depending on its function (amplification of DNA or expression of DNA) and the host cell for which it is compatible. The vector components generally include, but are not limited to, one or more of the following: a signal sequence, an origin of replication, one or more
15 marker genes, an enhancer element, a promoter, and a transcription termination sequence.

a. Signal Sequence Component

In general, a signal sequence may be a component of
20 the vector, or it may be a part of C-C CKR-1 DNA that is inserted into the vector.

b. Origin of Replication Component

Both expression and cloning vectors contain a nucleic
25 acid sequence that enables the vector to replicate in one or more selected host cells. Generally, in cloning vectors this sequence is one that enables the vector to replicate independently of the host chromosomal DNA, and includes origins of replication or autonomously replicating
30 sequences. Such sequences are well known for a variety of bacteria, yeast, and viruses. The origin of replication from the plasmid pBR322 is suitable for most Gram-negative bacteria, the 2 μ plasmid origin is suitable for yeast, and various viral origins (SV40, polyoma, adenovirus, VSV or
35 BPV) are useful for cloning vectors in mammalian cells. Generally, the origin of replication component is not needed for mammalian expression vectors (the SV40 origin may typically be used only because it contains the early promoter).

Most expression vectors are "shuttle" vectors, i.e. they are capable of replication in at least one class of organisms but can be transfected into another organism for expression. For example, a vector is cloned in *E. coli* and then the same vector is transfected into yeast or mammalian cells for expression even though it is not capable of replicating independently of the host cell chromosome.

DNA may also be amplified by insertion into the host genome. This is readily accomplished using *Bacillus* species as hosts, for example, by including in the vector a DNA sequence that is complementary to a sequence found in *Bacillus* genomic DNA. Transfection of *Bacillus* with this vector results in homologous recombination with the genome and insertion of C-C CKR-1 DNA. However, the recovery of genomic DNA encoding C-C CKR-1 is more complex than that of an exogenously replicated vector because restriction enzyme digestion is required to excise C-C CKR-1 DNA.

c. Selection Gene Component

Expression and cloning vectors should contain a selection gene, also termed a selectable marker. This gene encodes a protein necessary for the survival or growth of transformed host cells grown in a selective culture medium. Host cells not transformed with the vector containing the selection gene will not survive in the culture medium. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g. ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g. the gene encoding D-alanine racemase for *Bacilli*.

One example of a selection scheme utilizes a drug to arrest growth of a host cell. Those cells that are successfully transformed with a heterologous gene express a protein conferring drug resistance and thus survive the selection regimen. Examples of such dominant selection use the drugs neomycin (Southern et al., J. Molec. Appl. Genet. 1:327-341, 1982), mycophenolic acid (Mulligan et al.,

Science 209:1422-1427, 1980) or hygromycin (Sugden et al.,
Mol. Cell. Biol. 5:410-413, 1985). The three examples
given above employ bacterial genes under eukaryotic control
to convey resistance to the appropriate drug G418 or
5 neomycin (geneticin), xgpt (mycophenolic acid), or
hygromycin, respectively.

Another example of suitable selectable markers for
mammalian cells are those that enable the identification of
cells competent to take up C-C CKR-1 nucleic acid, such as
10 dihydrofolate reductase (DHFR) or thymidine kinase. The
mammalian cell transformants are placed under selection
pressure which only the transformants are uniquely adapted
to survive by virtue of having taken up the marker.
Selection pressure is imposed by culturing the
15 transformants under conditions in which the concentration
of selection agent in the medium is successively changed,
thereby leading to amplification of both the selection gene
and the DNA that encodes C-C CKR-1. Amplification is the
process by which genes in greater demand for the production
20 of a protein critical for growth are reiterated in tandem
within the chromosomes of successive generations of
recombinant cells. Increased quantities of C-C CKR-1 are
synthesized from the amplified DNA.

For example, cells transformed with the DHFR selection
25 gene are first identified by culturing all of the
transformants in a culture medium that contains
methotrexate (Mtx), a competitive antagonist of DHFR. An
appropriate host cell when wild-type DHFR is employed is
the Chinese hamster ovary (CHO) cell line deficient in DHFR
30 activity, prepared and propagated as described by Urlaub
and Chasin, Proc. Natl. Acad. Sci. U.S.A., 77(7):4216-4220,
1980). The transformed cells are then exposed to increased
levels of methotrexate. This leads to the synthesis of
multiple copies of the DHFR gene, and, concomitantly,
35 multiple copies of other DNA comprising the expression
vectors, such as the DNA encoding C-C CKR-1. This
amplification technique can be used with any otherwise
suitable host, e.g., ATCC No. CCL61 CHO-K1, notwithstanding
the presence of endogenous DHFR if, for example, a mutant

DHFR gene that is highly resistant to Mtx is employed (EP 117,060). Alternatively, host cells (particularly wild-type hosts that contain endogenous DHFR) transformed or co-transformed with DNA sequences encoding C-C CKR-1, wild-type DHFR protein, and another selectable marker such as aminoglycoside 3' phosphotransferase (APH) can be selected by cell growth in medium containing a selection agent for the selectable marker such as an aminoglycosidic antibiotic, e.g., kanamycin, neomycin, or G418.

10 A suitable selection gene for use in yeast is the *trp1* gene present in the yeast plasmid YRp7 (Stinchcomb et al., Nature 282:39-43, 1979); Kingsman et al., Gene 7:141-152, 1979); or Tschemper et al., Gene 10:157-166, 1980). The *trp1* gene provides a selection marker for a mutant strain
15 of yeast lacking the ability to grow in tryptophan, for example, ATCC No. 44076. The presence of the *trp1* lesion in the yeast host cell genome then provides an effective environment for detecting transformation by growth in the absence of tryptophan. Similarly, *Leu2*-deficient yeast
20 strains (ATCC 20,622 or 38,626) are complemented by known plasmids bearing the *Leu2* gene.

e. Promoter Component

Expression vectors usually contain a promoter that is
25 recognized by the host organism and is operably linked to C-C CKR-1 nucleic acid. Promoters are untranslated sequences located upstream (5') to the start codon of a structural gene (generally within about 100 to 1000 bp) that control the transcription and translation of a
30 particular nucleic acid sequence, such as C-C CKR-1, to which they are operably linked. Such promoters typically fall into two classes, inducible and constitutive. Inducible promoters are promoters that initiate increased levels of transcription from DNA under their control in
35 response to some change in culture conditions, e.g. the presence or absence of a nutrient or a change in temperature. At this time a large number of promoters recognized by a variety of potential host cells are well known. These promoters are operably linked to DNA encoding

C-C CKR-1 by removing the promoter from the source DNA by restriction enzyme digestion and inserting the isolated promoter sequence into the vector. Both the native C-C CKR-1 promoter sequence and many heterologous promoters may be used to direct amplification and/or expression of C-C CKR-1 DNA. However, heterologous promoters are preferred, as they generally permit greater transcription and higher yields of expressed C-C CKR-1 as compared to the native C-C CKR-1 promoter.

Promoters suitable for use with prokaryotic hosts include the β -lactamase and lactose promoter systems (Chang et al., Nature 275:617-624, 1978); and Goeddel et al., Nature 281:544-548, 1979), alkaline phosphatase, a tryptophan (trp) promoter system (Goeddel, Nucleic Acids Res. 8(18):4057-4074, 1980) and EP 36,776) and hybrid promoters such as the tac promoter (deBoer et al., Proc. Natl. Acad. Sci. U.S.A. 80:21-25, 1983). However, other known bacterial promoters are suitable. Their nucleotide sequences have been published, thereby enabling a skilled worker to operably ligate them to DNA encoding C-C CKR-1 (Siebenlist et al., Cell 20:269-281, 1980) using linkers or adapters to supply any required restriction sites. Promoters for use in bacterial systems also generally will contain a Shine-Dalgarno (S.D.) sequence operably linked to the DNA encoding C-C CKR-1.

Suitable promoting sequences for use with yeast hosts include the promoters for 3-phosphoglycerate kinase (Hitzeman et al., J. Biol. Chem. 255(24):12073-80, 1980) or other glycolytic enzymes (Hess et al., J. Adv. Enzyme Reg. 7:149-67, 1968); and Holland, Biochemistry 17:4900-4907, 1978), such as enolase, glyceraldehyde-3-phosphate dehydrogenase, hexokinase, pyruvate decarboxylase, phosphofructokinase, glucose-6-phosphate isomerase, 3-phosphoglycerate mutase, pyruvate kinase, triosephosphate isomerase, phosphoglucose isomerase, and glucokinase.

Other yeast promoters, which are inducible promoters having the additional advantage of transcription controlled by growth conditions, are the promoter regions for alcohol dehydrogenase 2, isocytochrome C, acid phosphatase,

degradative enzymes associated with nitrogen metabolism, metallothionein, glyceraldehyde-3-phosphate dehydrogenase, and enzymes responsible for maltose and galactose utilization. Suitable vectors and promoters for use in yeast expression are further described in Hitzeman et al., EP 73,657A. Yeast enhancers also are advantageously used with yeast promoters.

Promoter sequences are known for eukaryotes. Virtually all eukaryotic genes have an AT-rich region located approximately 25 to 30 bases upstream from the site where transcription is initiated. Another sequence found 70 to 80 bases upstream from the start of transcription of many genes is a CXCAAT region where X may be any nucleotide. At the 3' end of most eukaryotic genes is an AATAAA sequence that may be the signal for addition of the poly A tail to the 3' end of the coding sequence. All of these sequences are suitably inserted into mammalian expression vectors.

C-C CKR-1 transcription from vectors in mammalian host cells is controlled by promoters obtained from the genomes of viruses such as polyoma virus, fowlpox virus (UK 2,211,504 published 5 July 1989), adenovirus (such as Adenovirus 2), bovine papilloma virus, avian sarcoma virus, cytomegalovirus, a retrovirus, hepatitis B virus and most preferably Simian Virus 40 (SV40), from heterologous mammalian promoters, e.g. the actin promoter or an immunoglobulin promoter, from heat-shock promoters, and from the promoter normally associated with C-C CKR-1 sequence, provided such promoters are compatible with the host cell systems.

The early and late promoters of the SV40 virus are conveniently obtained as an SV40 restriction fragment that also contains the SV40 viral origin of replication (Fiers et al., Nature 273:113-120, 1978; Mulligan and Berg, Science 209:1422-1427, 1980; Pavlakis et al., Proc. Natl. Acad. Sci. U.S.A. 78:7398-7402, 1981). The immediate early promoter of the human cytomegalovirus is conveniently obtained as a HindIII E restriction fragment (Greenaway et al., Gene 18:355-360, 1982). A system for expressing DNA in

mammalian hosts using the bovine papilloma virus as a vector is disclosed in U.S. 4,419,446. A modification of this system is described in U.S. 4,601,978. See also Gray et al., Nature 295:503-508, 1982, on expressing cDNA

5 encoding immune interferon in monkey cells; Reyes et al., Nature 297:598-601, 1982, on expression of human β -interferon cDNA in mouse cells under the control of a thymidine kinase promoter from herpes simplex virus, Canaani and Berg, Proc. Natl. Acad. Sci. U.S.A. 79:5166-

10 5170, 1982, on expression of the human interferon β 1 gene in cultured mouse and rabbit cells, and Gorman et al., Proc. Natl. Acad. Sci. U.S.A. 79:6777-6781, 1982, on expression of bacterial CAT sequences in CV-1 monkey kidney cells, chicken embryo fibroblasts, Chinese hamster ovary

15 cells, HeLa cells, and mouse NIH-3T3 cells using the Rous sarcoma virus long terminal repeat as a promoter.

f. Enhancer Element Component

Transcription of a DNA encoding C-C CKR-1 of this

20 invention by higher eukaryotes is often increased by inserting an enhancer sequence into the vector. Enhancers are cis-acting elements of DNA, usually about from 10-300 bp, that act on a promoter to increase its transcription. Enhancers are relatively orientation and position

25 independent having been found 5' (Laimins et al., Proc. Natl. Acad. Sci. U.S.A. 78:464-8, 1981) and 3' (Lusky et al., Mol. Cell Bio. 3(6):1108-1122, 1983) to the transcription unit, within an intron (Banerji et al., Cell 33:729-740, 1983) as well as within the coding sequence

30 itself (Osborne et al., Mol. Cell Bio. 4(7):1293-1305, 1984). Many enhancer sequences are now known from mammalian genes (globin, elastase, albumin, α -fetoprotein and insulin). Typically, however, one will use an enhancer from a eukaryotic cell virus. Examples include the SV40

35 enhancer on the late side of the replication origin (bp 100-270), the cytomegalovirus early promoter enhancer, the polyoma enhancer on the late side of the replication origin, and adenovirus enhancers. See also Yaniv, Nature 297:17-18, 1982, on enhancing elements for activation of

eukaryotic promoters. The enhancer may be spliced into the vector at a position 5' or 3' to C-C CKR-1 DNA, but is preferably located at a site 5' from the promoter.

5 g. Transcription Termination Component

Expression vectors used in eukaryotic host cells (yeast, fungi, insect, plant, animal, human, or nucleated cells from other multicellular organisms) will also contain sequences necessary for the termination of transcription
10 and for stabilizing the mRNA. Such sequences are commonly available from the 5' and, occasionally 3' untranslated regions of eukaryotic or viral DNAs or cDNAs. These regions contain nucleotide segments transcribed as polyadenylated fragments in the untranslated portion of the
15 mRNA encoding C-C CKR-1. The 3' untranslated regions also include transcription termination sites.

Suitable vectors containing one or more of the above listed components and the desired coding and control sequences are constructed by standard ligation techniques.
20 Isolated plasmids or DNA fragments are cleaved, tailored, and religated in the form desired to generate the plasmids required.

For analysis to confirm correct sequences in plasmids constructed, the ligation mixtures are used to transform E.
25 coli K12 strain 294 (ATCC 31,446) and successful transformants selected by ampicillin or tetracycline resistance where appropriate. Plasmids from the transformants are prepared, analyzed by restriction endonuclease digestion, and/or sequenced by the method of
30 Messing et al., Nucleic Acids Res. 9(2):309-321, 1981, or by the method of Maxam et al., Methods in Enzymology 65:499-560, 1980.

Particularly useful in the practice of this invention are expression vectors that provide for the transient
35 expression in mammalian cells of DNA encoding C-C CKR-1. In general, transient expression involves the use of an expression vector that is able to replicate efficiently in a host cell, such that the host cell accumulates many copies of the expression vector and, in turn, synthesizes

high levels of a desired polypeptide encoded by the expression vector. Transient expression systems, comprising a suitable expression vector and a host cell, allow for the convenient positive identification of polypeptides encoded by cloned DNAs, as well as for the rapid screening of such polypeptides for desired biological or physiological properties. Thus, transient expression systems are particularly useful in the invention for purposes of identifying analogs and variants of C-C CKR-1 that have C-C CKR-1-like activity, and for analysis of the effect of the binding of chemokine variants to C-C CKR-1.

Other methods, vectors, and host cells suitable for adaptation to the synthesis of C-C CKR-1 in recombinant vertebrate cell culture are described in Gething et al., Nature 293:620-625, 1981; Mantei et al., Nature 281:40-46, 1979); EP 117,060; and EP 117,058. A particularly useful plasmid for mammalian cell culture expression of C-C CKR-1 is pRK5 (EP pub. no. 307,247) or pSVI6B (U.S. Ser. No. 07/441,574 filed 22 November 1989).

20

4. Selection and Transformation of Host Cells

Suitable host cells for cloning or expressing C-C CKR-1 expression vectors are the prokaryote, yeast, or higher eukaryotic cells described above. Suitable prokaryotes include eubacteria, such as Gram-negative or Gram-positive organisms, for example, *E. coli*, *Bacilli* such as *B. subtilis*, *Pseudomonas* species such as *P. aeruginosa*, *Salmonella typhimurium*, or *Serratia marcescens*. One preferred *E. coli* cloning host is *E. coli* 294 (ATCC 31,446), although other strains such as *E. coli* B, *E. coli* χ 1776 (ATCC 31,537), and *E. coli* W3110 (ATCC 27,325) are suitable. These examples are illustrative rather than limiting. Preferably the host cell should secrete minimal amounts of proteolytic enzymes. Alternatively, in vitro methods of cloning, e.g. PCR or other nucleic acid polymerase reactions, are suitable.

In addition to prokaryotes, eukaryotic microbes such as filamentous fungi or yeast are suitable hosts for vectors containing C-C CKR-1 DNA. *Saccharomyces*

- cerevisiae, or common baker's yeast, is the most commonly used among lower eukaryotic host microorganisms. However, a number of other genera, species, and strains are commonly available and useful to practice the invention, such as S.
- 5 pombe (Beach and Nurse, Nature 290:140-143, 1981), Kluyveromyces lactis (Louvencourt et al., J. Bacteriol. 154(2):737-742, 1983), Pichia pastoris (EP 183,070), Trichoderma reesia (EP 244,234), Neurospora crassa (Case et al., Proc. Natl. Acad. Sci. U.S.A. 76:5259-5263, 1979), and
- 10 Aspergillus hosts such as A. nidulans (Ballance et al., Biochem. Biophys. Res. Commun. 112:284-289, 1983); Tilburn et al., Gene 26:205-221, 1983); Yelton et al., Proc. Natl. Acad. Sci. U.S.A. 81:1470-1474, 1984) and A. niger (Kelly and Hynes, EMBO J. 4:475-479, 1985).
- 15 Suitable host cells for the expression of glycosylated C-C CKR-1 polypeptide are derived from multicellular organisms. Such host cells are capable of complex processing and glycosylation activities. In principle, any higher eukaryotic cell culture is workable, whether from
- 20 vertebrate or invertebrate culture. Examples of invertebrate cells include plant and insect cells. Numerous baculoviral strains and variants and corresponding permissive insect host cells from hosts such as Spodoptera frugiperda (caterpillar), Aedes aegypti (mosquito), Aedes
- 25 albopictus (mosquito), Drosophila melanogaster (fruit fly), and Bombyx mori host cells have been identified. See, e.g., Luckow et al., Bio/Technology 6:47-55, 1988); Miller et al., in Genetic Engineering, Setlow, J.K. et al., 8:277-279 (Plenum Publishing, 1986), and Maeda et al., Nature
- 30 315:592-594, 1985). A variety of such viral strains are publicly available, e.g., the L-1 variant of Autographa californica NPV and the Bm-5 strain of Bombyx mori NPV, and such viruses may be used as the virus according to the present invention, particularly for transfection of
- 35 Spodoptera frugiperda cells.

Plant cell cultures of cotton, corn, potato, soybean, petunia, tomato, and tobacco can be utilized as hosts. Typically, plant cells are transfected by incubation with certain strains of the bacterium Agrobacterium tumefaciens,

which has been previously manipulated to contain C-C CKR-1 DNA. During incubation of the plant cell culture with *A. tumefaciens*, the DNA encoding C-C CKR-1 is transferred to the plant cell host such that it is transfected, and will, under appropriate conditions, express C-C CKR-1 DNA. In addition, regulatory and signal sequences compatible with plant cells are available, such as the nopaline synthase promoter and polyadenylation signal sequences. Depicker et al., J. Mol. Appl. Gen. 1: 561-573, 1982). In addition, DNA segments isolated from the upstream region of the T-DNA 780 gene are capable of activating or increasing transcription levels of plant-expressible genes in recombinant DNA-containing plant tissue. See EP 321,196 published 21 June 1989.

However, interest has been greatest in vertebrate cells, and propagation of vertebrate cells in culture (tissue culture) has become a routine procedure in recent years (Tissue Culture, Academic Press, Kruse and Patterson, eds., 1973). Examples of useful mammalian host cell lines are monkey kidney CV1 line transformed by SV40 (COS-7, ATCC CRL 1651); human embryonic kidney line (293 or 293 cells subcloned for growth in suspension culture, Graham et al., J. Gen. Virol. 36:59-72, 1977); baby hamster kidney cells (BHK, ATCC CCL 10); Chinese hamster ovary cells/-DHFR (CHO, Urlaub and Chasin, Proc. Natl. Acad. Sci. U.S.A. 77(7):4216-4220, 1980); mouse sertoli cells (TM4, Mather, Biol. Reprod. 23:243-251, 1980); monkey kidney cells (CV1 ATCC CCL 70); African green monkey kidney cells (VERO-76, ATCC CRL-1587); human cervical carcinoma cells (HeLa, ATCC CCL 2); canine kidney cells (MDCK, ATCC CCL 34); buffalo rat liver cells (BRL 3A, ATCC CRL 1442); human lung cells (W138, ATCC CCL 75); human liver cells (Hep G2, HB 8065); mouse mammary tumor (MMT 060562, ATCC CCL51); TRI cells (Mather et al., Annals N.Y. Acad. Sci. 383:44-68, 1982); MRC 5 cells; FS4 cells; and a human hepatoma cell line (Hep G2). Preferred host cells are human embryonic kidney 293 and Chinese hamster ovary cells.

The host chosen for expression may also be a multicellular organism, as in a transgenic animal. Such

animals have been produced by transfection of germ cells, somatic cells, or embryos with heterologous DNA, suitably implanting the transfected cells and allowing the cells to mature into or stably integrate into adult animals

5 containing the heterologous DNA. A reproducible percentage of such animals transcribe and express the heterologous DNA as protein which can be identified in tissues including blood or serum. Suitable methods for making transgenic animals are described in U.S. Patent 4,396,601 and Palmiter
10 et al., Nature 300:611-615, 1982.

Host cells are transfected and preferably transformed with the above-described expression or cloning vectors of this invention and cultured in conventional nutrient media modified as appropriate for inducing promoters, selecting
15 transformants, or amplifying the genes encoding the desired sequences.

Transfection refers to the taking up of an expression vector by a host cell whether or not any coding sequences are in fact expressed. Numerous methods of transfection
20 are known to the ordinarily skilled artisan, for example, CaPO_4 and electroporation. Successful transfection is generally recognized when any indication of the operation of this vector occurs within the host cell.

Transformation means introducing DNA into an organism
25 so that the DNA is replicable, either as an extrachromosomal element or by chromosomal integrant. Depending on the host cell used, transformation is done using standard techniques appropriate to such cells. The calcium treatment employing calcium chloride, as described
30 in section 1.82 of Sambrook et al., is generally used for prokaryotes or other cells that contain substantial cell-wall barriers. Infection with *Agrobacterium tumefaciens* is used for transformation of certain plant cells, as described by Shaw et al., Gene 23:315-330, 1983) and WO
35 89/05859, published 29 June 1989. For mammalian cells without such cell walls, the calcium phosphate precipitation method described in sections 16.30-16.37 of Sambrook et al., supra, is preferred. General aspects of mammalian cell host system transformations have been

described by Axel in U.S. 4,399,216, issued 16 August 1983. Transformations into yeast are typically carried out according to the method of Van Solingen et al., J. Bacteriol. 130(2):946-947, 1977) and Hsiao et al., Proc. Natl. Acad. Sci. U.S.A., 76(8):3829-3833, 1979). However, other methods for introducing DNA into cells such as by nuclear injection, electroporation, or by protoplast fusion may also be used.

10 5. Culturing the Host Cells

Prokaryotic cells used to produce C-C CKR-1 polypeptide of this invention are cultured in suitable media as described generally in Sambrook et al., supra.

15 The mammalian host cells used to produce C-C CKR-1 of this invention may be cultured in a variety of media. Commercially available media such as Ham's F10 (Sigma), Minimal Essential Medium (MEM, Sigma), RPMI-1640 (Sigma), and Dulbecco's Modified Eagle's Medium (DMEM, Sigma) are
20 suitable for culturing the host cells. In addition, any of the media described in Ham and McKeehan, Meth. Enz. 58:44-93, 1979, Barnes and Sato, Anal. Biochem. 102:255-270, 1980, U.S. 4,767,704; 4,657,866; 4,927,762; or 4,560,655; WO 90/03430; WO 87/00195; U.S. Pat. Re. 30,985; or U.S.
25 5,122,469 may be used as culture media for the host cells. Any of these media may be supplemented as necessary with hormones and/or other growth factors (such as insulin, transferrin, or epidermal growth factor), salts (such as sodium chloride, calcium, magnesium, and phosphate),
30 buffers (such as HEPES), nucleosides (such as adenosine and thymidine), antibiotics (such as GentamycinTM drug), trace elements (defined as inorganic compounds usually present at final concentrations in the micromolar range), and glucose or an equivalent energy source. Any other necessary
35 supplements may also be included at appropriate concentrations that would be known to those skilled in the art. The culture conditions, such as temperature, pH, and the like, are those previously used with the host cell selected for expression, and will be apparent to the

ordinarily skilled artisan.

The host cells referred to in this disclosure encompass cells in in vitro culture as well as cells that are within a host animal.

5

6. Detecting Gene Amplification/Expression

Gene amplification and/or expression may be measured in a sample directly, for example, by conventional Southern blotting, northern blotting to quantitate the transcription of mRNA (Thomas, Proc. Natl. Acad. Sci. U.S.A. 10 77:5201-5205, 1980), dot blotting (DNA analysis), or in situ hybridization, using an appropriately labeled probe. Various labels may be employed, most commonly radioisotopes, particularly ^{32}P . However, other techniques 15 may also be employed, such as using biotin-modified nucleotides for introduction into a polynucleotide. The biotin then serves as the site for binding to avidin or antibodies, which may be labeled with a wide variety of labels, such as radionuclides, fluorescers, enzymes, or the 20 like.

Alternatively, antibodies may be employed that can recognize specific duplexes, including DNA duplexes, RNA duplexes, and DNA-RNA hybrid duplexes or DNA-protein duplexes. The antibodies in turn may be labeled and the 25 assay may be carried out where the duplex is bound to a surface, so that upon the formation of duplex on the surface, the presence of antibody bound to the duplex can be detected.

Gene expression, alternatively, may be measured by 30 immunological methods, such as immunohistochemical staining of tissue sections and assay of cell culture or body fluids, to quantitate directly the expression of gene product. With immunohistochemical staining techniques, a cell sample is prepared, typically by dehydration and 35 fixation, followed by reaction with labeled antibodies specific for the gene product coupled, where the labels are usually visually detectable, such as enzymatic labels, fluorescent labels, luminescent labels, and the like. A particularly sensitive staining technique suitable for use

in the present invention is described by Hsu et al.,
Am. J. Clin. Path. 75:734-738, 1980.

Antibodies useful for immunohistochemical staining
and/or assay of sample fluids may be either monoclonal or
5 polyclonal, and may be prepared in any mammal.
Conveniently, the antibodies may be prepared against a
native or synthetic C-C CKR-1 polypeptide or variant
thereof.

10 7. Purification of C-C CKR-1 Polypeptide

C-C CKR-1 is recovered from cell cultures by
solubilizing cell membranes in detergent.

When a human C-C CKR-1 is expressed in a recombinant
cell other than one of human origin, C-C CKR-1 is
15 completely free of proteins or polypeptides of human
origin. However, it is necessary to purify C-C CKR-1 from
recombinant cell proteins or polypeptides to obtain
preparations that are substantially homogeneous by protein
as to C-C CKR-1. As a first step, the cells are
20 centrifuged to separate them from culture medium, followed
by suitable purification procedures such as: fractionation
on immunoaffinity or ion-exchange columns; ethanol
precipitation; reverse phase HPLC; chromatography on silica
or on a cation exchange resin such as DEAE;
25 chromatofocusing; SDS-PAGE; ammonium sulfate precipitation;
gel filtration using, for example, Sephadex G-75.

C-C CKR-1 variants in which residues have been
deleted, inserted or substituted are recovered in the same
fashion as the native C-C CKR-1, taking account of any
30 substantial changes in properties occasioned by the
variation. For example, preparation of a C-C CKR-1 fusion
with another protein or polypeptide, e.g. a bacterial or
viral antigen, facilitates purification; an immunoaffinity
column containing antibody to the antigen can be used to
35 adsorb the fusion. Immunoaffinity columns such as a rabbit
polyclonal anti-C-C CKR-1 column can be employed to absorb
C-C CKR-1 variant by binding it to at least one remaining
immune epitope. A protease inhibitor such as phenyl methyl
sulfonyl fluoride (PMSF) also may be useful to inhibit

proteolytic degradation during purification, and antibiotics may be included to prevent the growth of adventitious contaminants. One skilled in the art will appreciate that purification methods suitable for native C-
5 C CKR-1 may require modification to account for changes in the character of C-C CKR-1 or its variants upon expression in recombinant cell culture.

8. Covalent Modifications of C-C CKR-1 Polypeptides

10 Covalent modifications of C-C CKR-1 polypeptide or its glycosyl substituents are included within the scope of this invention. Both native C-C CKR-1 and amino acid sequence variants of C-C CKR-1 may be covalently modified. Covalent
15 modifications of C-C CKR-1, fragments thereof or antibodies thereto are introduced into the molecule by reacting targeted amino acid residues of C-C CKR-1, fragments
thereof, or C-C CKR-1 antibody with an organic derivatizing agent that is capable of reacting with selected side chains or the N- or C-terminal residues. Most commonly, C-C CKR-1
20 and its antibodies are covalently bonded to detectable groups used in diagnosis, e.g. enzymes, radio isotopes, spin labels, antigens, fluorescent or chemiluminescent groups and the like.

Cysteinylnyl residues most commonly are reacted with α -
25 haloacetates (and corresponding amines), such as chloroacetic acid or chloroacetamide, to give carboxymethyl or carboxyamidomethyl derivatives. Cysteinylnyl residues also are derivatized by reaction with bromotrifluoroacetone, α -bromo- β -(5-imidazole)propionic acid, chloroacetyl
30 phosphate, N-alkylmaleimides, 3-nitro-2-pyridyl disulfide, methyl 2-pyridyl disulfide, p-chloromercuribenzoate, 2-chloromercuri-4-nitrophenol, or chloro-7-nitrobenzo-2-oxa-1,3-diazole.

Histidyl residues are derivatized by reaction with
35 diethylpyrocarbonate at pH 5.5-7.0 because this agent is relatively specific for the histidyl side chain. Para-bromophenacyl bromide also is useful; the reaction is preferably performed in 0.1 M sodium cacodylate at pH 6.0.

Lysinylnyl and amino terminal residues are reacted with

succinic or other carboxylic acid anhydrides.

Derivatization with these agents has the effect of reversing the charge of the lysinyl residues. Other suitable reagents for derivatizing α -amino-containing

5 residues include imidoesters such as methyl picolinimate; pyridoxal phosphate; pyridoxal; chloroborohydride; trinitrobenzenesulfonic acid; O-methylisourea; 2,4-pentanedione; and transaminase-catalyzed reaction with glyoxylate.

10 Arginyl residues are modified by reaction with one or several conventional reagents, among them phenylglyoxal, 2,3-butanedione, 1,2-cyclohexanedione, and ninhydrin. Derivatization of arginine residues requires that the reaction be performed in alkaline conditions because of the
15 high pK_a of the guanidine functional group. Furthermore, these reagents may react with the groups of lysine as well as the arginine epsilon-amino group.

The specific modification of tyrosyl residues may be made, with particular interest in introducing spectral
20 labels into tyrosyl residues by reaction with aromatic diazonium compounds or tetranitromethane. Most commonly, N-acetylimidazole and tetranitromethane are used to form O-acetyl tyrosyl species and 3-nitro derivatives, respectively. Tyrosyl residues are iodinated using ^{125}I or
25 ^{131}I to prepare labeled proteins for use in radioimmunoassay, the chloramine T method described above being suitable.

Carboxyl side groups (aspartyl or glutamyl) are selectively modified by reaction with carbodiimides ($\text{R}'\text{-N}=\text{C}=\text{N-R}'$), where R and R' are different alkyl groups, such
30 as 1-cyclohexyl-3-(2-morpholinyl-4-ethyl) carbodiimide or 1-ethyl-3-(4-azonia-4,4-dimethylpentyl) carbodiimide. Furthermore, aspartyl and glutamyl residues are converted to asparaginyl and glutaminyl residues by reaction with
35 ammonium ions.

Derivatization with bifunctional agents is useful for cross-linking C-C CKR-1, its fragments or antibodies to a water-insoluble support matrix or surface for use in methods for purifying anti-C-C CKR-1 antibodies, and vice

versa. Commonly used cross-linking agents include, e.g., 1,1-bis(diazoacetyl)-2-phenylethane, glutaraldehyde, N-hydroxysuccinimide esters, for example, esters with 4-azidosalicylic acid, homobifunctional imidoesters, including disuccinimidyl esters such as 3,3'-dithiobis-(succinimidylpropionate), and bifunctional maleimides such as bis-N-maleimido-1,8-octane. Derivatizing agents such as methyl-3-[(p-azidophenyl)dithio]propioimide yield photoactivatable intermediates that are capable of forming cross-links in the presence of light. Alternatively, reactive water-insoluble matrices such as cyanogen bromide-activated carbohydrates and the reactive substrates described in U.S. 3,969,287; 3,691,016; 4,195,128; 4,247,642; 4,229,537; and 4,330,440 are employed for protein immobilization.

Glutamyl and asparaginyl residues are frequently deamidated to the corresponding glutamyl and aspartyl residues, respectively. Alternatively, these residues are deamidated under mildly acidic conditions. Either form of these residues falls within the scope of this invention.

Other modifications include hydroxylation of proline and lysine, phosphorylation of hydroxyl groups of seryl or threonyl residues, methylation of the α -amino groups of lysine, arginine, and histidine side chains (T.E. Creighton, Proteins: Structure and Molecular Properties, W.H. Freeman & Co., San Francisco, pp. 79-86 1983), acetylation of the N-terminal amine, and amidation of any C-terminal carboxyl group.

Another type of covalent modification of C-C CKR-1 polypeptide included within the scope of this invention comprises altering the native glycosylation pattern of the polypeptide. By altering is meant deleting one or more carbohydrate moieties found in the native polypeptide, and/or adding one or more glycosylation sites that are not present in the native polypeptide.

Glycosylation of polypeptides is typically either N-linked or O-linked. N-linked refers to the attachment of the carbohydrate moiety to the side chain of an asparagine residue. The tri-peptide sequences asparagine-X-serine and

asparagine-X-threonine, where X is any amino acid except proline, are the recognition sequences for enzymatic attachment of the carbohydrate moiety to the asparagine side chain. Thus, the presence of either of these tri-peptide sequences in a polypeptide creates a potential glycosylation site. O-linked glycosylation refers to the attachment of one of the sugars N-acetylgalactosamine, galactose, or xylose, to a hydroxyamino acid, most commonly serine or threonine, although 5-hydroxyproline or 5-hydroxylysine may also be used.

Addition of glycosylation sites to C-C CKR-1 polypeptide is conveniently accomplished by altering the amino acid sequence such that it contains one or more of the above-described tri-peptide sequences (for N-linked glycosylation sites). The alteration may also be made by the addition of, or substitution by, one or more serine or threonine residues to the native C-C CKR-1 sequence (for O-linked glycosylation sites). For ease, C-C CKR-1 amino acid sequence is preferably altered through changes at the DNA level, particularly by mutating the DNA encoding C-C CKR-1 polypeptide at preselected bases such that codons are generated that will translate into the desired amino acids. The DNA mutation(s) may be made using methods described above under the heading of "Amino Acid Sequence Variants of C-C CKR-1 Polypeptide".

Another means of increasing the number of carbohydrate moieties on C-C CKR-1 polypeptide is by chemical or enzymatic coupling of glycosides to the polypeptide. These procedures are advantageous in that they do not require production of the polypeptide in a host cell that has glycosylation capabilities for N- and O-linked glycosylation. Depending on the coupling mode used, the sugar(s) may be attached to (a) arginine and histidine, (b) free carboxyl groups, (c) free sulfhydryl groups such as those of cysteine, (d) free hydroxyl groups such as those of serine, threonine, or hydroxyproline, (e) aromatic residues such as those of phenylalanine, tyrosine, or tryptophan, or (f) the amide group of glutamine. These methods are described in WO 87/05330 published 11 September

1987, and in Aplin and Wriston (CRC Crit. Rev. Biochem. pp. 259-306, 1981).

Removal of carbohydrate moieties present on the native C-C CKR-1 polypeptide may be accomplished chemically or enzymatically. Chemical deglycosylation requires exposure of the polypeptide to the compound trifluoromethanesulfonic acid, or an equivalent compound. This treatment results in the cleavage of most or all sugars except the linking sugar (N-acetylglucosamine or N-acetylgalactosamine), while leaving the polypeptide intact. Chemical deglycosylation is described by Hakimuddin et al. (Arch. Biochem. Biophys. 259:52-57, 1987) and by Edge et al. (Anal. Biochem. 118:131-137, 1981). Enzymatic cleavage of carbohydrate moieties on polypeptides can be achieved by the use of a variety of endo- and exo- glycosidases as described by Thotakura et al. (Meth. Enzymol. 138:350-359, 1987).

Glycosylation at potential glycosylation sites may be prevented by the use of the compound tunicamycin as described by Duskin et al. (J. Biol. Chem. 257:3105-3109, 1982). Tunicamycin blocks the formation of protein-N-glycoside linkages.

The C-C CKR-1 also may be entrapped in microcapsules prepared, for example, by coacervation techniques or by interfacial polymerization (for example, hydroxymethylcellulose or gelatin-microcapsules and poly-[methacrylate) microcapsules, respectively), in colloidal drug delivery systems (for example, liposomes, albumin microspheres, microemulsions, nano-particles and nanocapsules), or in macroemulsions. Such techniques are disclosed in Remington's Pharmaceutical Sciences, 16th edition, Osol, A., ed., 1980).

C-C CKR-1 preparations are also useful in generating antibodies, for use as standards in assays for C-C CKR-1 (e.g. by labeling C-C CKR-1 for use as a standard in a radioimmunoassay, enzyme-linked immunoassay, or radioreceptor assay), in affinity purification techniques, and in competitive-type receptor binding assays when labeled with radioiodine, enzymes, fluorophores, spin labels, and the like.

Since it is often difficult to predict in advance the characteristics of a variant C-C CKR-1, it will be appreciated that some screening of the recovered variant will be needed to select the optimal variant. For example, a change in the immunological character of C-C CKR-1 molecule, such as affinity for a given antibody, is measured by a competitive-type immunoassay. The variant is assayed for changes in the suppression or enhancement of its activity by comparison to the activity observed for native C-C CKR-1 in the same assay. Other potential modifications of protein or polypeptide properties such as redox or thermal stability, hydrophobicity, susceptibility to proteolytic degradation, or the tendency to aggregate with carriers or into multimers are assayed by methods well known in the art.

9. Therapeutic Compositions and Administration of C-C CKR-1

Therapeutic formulations of C-C CKR-1 (including its C-C CKR-1 binding fragments) or antibodies thereto are prepared for storage by mixing C-C CKR-1 having the desired degree of purity with optional physiologically acceptable carriers, excipients, or stabilizers (Remington's Pharmaceutical Sciences, supra), in the form of lyophilized cake or aqueous solutions. Acceptable carriers, excipients or stabilizers are nontoxic to recipients at the dosages and concentrations employed, and include buffers such as phosphate, citrate, and other organic acids; antioxidants including ascorbic acid; low molecular weight (less than about 10 residues) polypeptides; proteins, such as serum albumin, gelatin, or immunoglobulins; hydrophilic polymers such as polyvinylpyrrolidone; amino acids such as glycine, glutamine, asparagine, arginine or lysine; monosaccharides, disaccharides, and other carbohydrates including glucose, mannose, or dextrans; chelating agents such as EDTA; sugar alcohols such as mannitol or sorbitol; salt-forming counterions such as sodium; and/or nonionic surfactants such as Tween, Pluronic, or polyethylene glycol (PEG).

The C-C CKR-1 or antibody to be used for in vivo

administration must be sterile. This is readily accomplished by filtration through sterile filtration membranes, prior to or following lyophilization and reconstitution. The C-C CKR-1 ordinarily will be stored in
5 lyophilized form or in solution.

Therapeutic C-C CKR-1 or antibody compositions generally are placed into a container having a sterile access port, for example, an intravenous solution bag or vial having a stopper pierceable by a hypodermic injection
10 needle.

The route of C-C CKR-1 or antibody administration is in accord with known methods, e.g. injection or infusion by intravenous, intraperitoneal, intracerebral, intramuscular, intraocular, intraarterial, or intralesional routes, or by
15 sustained release systems as noted below.

Suitable examples of sustained-release preparations include semipermeable polymer matrices in the form of shaped articles, e.g. films, or microcapsules. Sustained release matrices include polyesters, hydrogels,
20 polylactides (U.S. 3,773,919, EP 58,481), copolymers of L-glutamic acid and γ ethyl-L-glutamate (Sidman et al., Biopolymers 22:547-556, 1983), poly (2-hydroxyethyl-methacrylate) (Langer et al., J. Biomed. Mater. Res. 15:167-277, 1981; Langer, Chem. Tech., 12:98-105, 1982),
25 ethylene vinyl acetate (Langer et al., supra) or poly-D-(-)-3-hydroxybutyric acid (EP 133,988). Sustained-release C-C CKR-1 or antibody compositions also include liposomally entrapped C-C CKR-1 or antibody. Liposomes containing C-C CKR-1 or antibody are prepared by methods known per se: DE
30 3,218,121; Epstein et al., Proc. Natl. Acad. Sci. U.S.A. 82:3688-3692, 1985; Hwang et al., Proc. Natl. Acad. Sci. U.S.A. 77:4030-4034, 1980; EP 52,322; EP 36,676; EP 88,046; EP 143,949; EP 142,641; Japanese patent application 83-118008; U.S. 4,485,045 and 4,544,545; and EP 102,324.
35 Ordinarily the liposomes are of the small (about 200-800 Angstroms) unilamellar type in which the lipid content is greater than about 30 mol. % cholesterol, the selected proportion being adjusted for the optimal C-C CKR-1 or antibody therapy.

An effective amount of C-C CKR-1 or antibody to be employed therapeutically will depend, for example, upon the therapeutic objectives, the route of administration, and the condition of the patient. For example, it is expected that C-C CKR-1 will be therapeutically effective in the treatment of cytokine-mediated inflammation. Accordingly, it will be necessary for the therapist to titer the dosage and modify the route of administration as required to obtain the optimal therapeutic effect. Typically, the clinician will administer C-C CKR-1 or antibody until a dosage is reached that achieves the desired effect. The progress of this therapy is easily monitored by conventional assays.

10. C-C CKR-1 Antibody Preparation

Polyclonal antibodies to C-C CKR-1 generally are raised in animals by multiple subcutaneous (sc) or intraperitoneal (ip) injections of C-C CKR-1 and an adjuvant. Immunization with recombinant cells transformed with C-C CKR-1 (e.g. mouse or CHO cells transformed with human C-C CKR-1) may be satisfactory, or it may be useful to separate C-C CKR-1 and conjugate it or a fragment containing the target amino acid sequence to a protein that is immunogenic in the species to be immunized, e.g., keyhole limpet hemocyanin, serum albumin, bovine thyroglobulin, or soybean trypsin inhibitor using a bifunctional or derivatizing agent, for example, maleimidobenzoyl sulfosuccinimide ester (conjugation through cysteine residues), N-hydroxysuccinimide (through lysine residues), glutaraldehyde, succinic anhydride, SOCl_2 , or $\text{R}^1\text{N} = \text{C} = \text{NR}$, where R and R^1 are different alkyl groups.

Animals ordinarily are immunized against the cells or immunogenic conjugates or derivatives by combining 1 mg or 1 μg of C-C CKR-1 in Freund's complete adjuvant and injecting the solution intradermally at multiple sites. One month later the animals are boosted with 1/5 to 1/10 the original amount of conjugate in Freund's complete adjuvant by subcutaneous injection at multiple sites. 7 to

14 days later animals are bled and the serum is assayed for anti-C-C CKR-1 titer. Animals are boosted until the titer plateaus. Preferably, the animal is boosted with the conjugate of the same C-C CKR-1, but conjugated to a different protein and/or through a different cross-linking agent. Conjugates also can be made in recombinant cell culture as protein fusions. Also, aggregating agents such as alum are used to enhance the immune response.

Another option is to employ combinatorial variable domain libraries and screening methods to identify the desired anti-C-C CKR-1 antibodies.

Monoclonal antibodies are prepared by recovering spleen cells from immunized animals and immortalizing the cells in conventional fashion, e.g. by fusion with myeloma cells or by Epstein-Barr (EB)-virus transformation and screening for clones expressing the desired antibody.

The monoclonal antibody preferably is specific for each target C-C CKR-1 polypeptide. The antibody is selected to be either agonistic, antagonistic or to have no effect on the activity of or binding of the C-C CKR-1.

11. Uses of C-C CKR-1 Nucleic Acid, and Antibodies

The nucleic acid encoding C-C CKR-1 may be used as a diagnostic for tissue specific typing. For example, such procedures as in situ hybridization, and northern and Southern blotting, and PCR analysis may be used to determine whether DNA and/or RNA encoding C-C CKR-1 are present in the cell type(s) being evaluated.

Isolated C-C CKR-1 polypeptide may be used in quantitative diagnostic assays as a standard or control against which samples e.g., from erythrocytes, containing unknown quantities of C-C CKR-1 may be compared. Recombinant cells which express C-C CKR-1 can be used in assays for C-C CKR-1 ligands in the same fashion as for example neutrophils are used in IL-8 assays. The C-C CKR-1 polypeptides, fragments or cells (as such, or derivatized) also can be used as immunogens in the production of antibodies to C-C CKR-1, or for the purification of such antibodies from ascites or recombinant cell culture media.

C-C CKR-1 antibodies are useful in diagnostic assays for C-C CKR-1 expression in specific cells or tissues wherein the antibodies are labeled in the same fashion as C-C CKR-1 described above and/or are immobilized on an insoluble matrix. C-C CKR-1 antibodies also are useful for the affinity purification of C-C CKR-1 from recombinant cell culture or natural sources. The C-C CKR-1 antibodies that do not detectably cross-react with other chemokine receptors can be used to purify each C-C CKR-1 free from other homologous chemokine receptors. C-C CKR-1 antibodies that are PF4 superfamily antagonists are useful as anti-inflammatory agents or in the treatment of other PF4 superfamily-mediated disorders.

Suitable diagnostic assays for C-C CKR-1 and its antibodies are well known per se. Such assays include competitive and sandwich assays, and steric inhibition assays. Competitive and sandwich methods employ a phase-separation step as an integral part of the method while steric inhibition assays are conducted in a single reaction mixture. Fundamentally, the same procedures are used for the assay of C-C CKR-1 and for substances that bind C-C CKR-1, although certain methods will be favored depending upon the molecular weight of the substance being assayed. Therefore, the substance to be tested is referred to as an analyte, irrespective of its status otherwise as an antigen or antibody, and proteins that bind to the analyte are denominated binding partners, whether they be antibodies, cell surface receptors, or antigens.

Analytical methods for C-C CKR-1 or its antibodies all use one or more of the following reagents: labeled analyte analog, immobilized analyte analog, labeled binding partner, immobilized binding partner and steric conjugates. The labeled reagents also are known as "tracers."

The label used (and this is also useful to label C-C CKR-1 nucleic acid for use as a probe) is any detectable functionality that does not interfere with the binding of analyte and its binding partner. Numerous labels are known for use in immunoassay, examples including moieties that may be detected directly, such as fluorochrome,

chemiluminescent, and radioactive labels, as well as moieties, such as enzymes, that must be reacted or derivatized to be detected. Examples of such labels include the radioisotopes ^{32}P , ^{14}C , ^{125}I , ^3H , and ^{131}I ,
5 fluorophores such as rare earth chelates or fluorescein and its derivatives, rhodamine and its derivatives, dansyl, umbelliferone, luciferases, e.g., firefly luciferase and bacterial luciferase (U.S. Pat. No. 4,737,456), luciferin, 2,3-dihydrophthalazinediones, horseradish peroxidase (HRP),
10 alkaline phosphatase, β -galactosidase, glucoamylase, lysozyme, saccharide oxidases, e.g., glucose oxidase, galactose oxidase, and glucose-6-phosphate dehydrogenase, heterocyclic oxidases such as uricase and xanthine oxidase, coupled with an enzyme that employs hydrogen peroxide to
15 oxidize a dye precursor such as HRP, lactoperoxidase, or microperoxidase, biotin/avidin, spin labels, bacteriophage labels, stable free radicals, and the like.

Conventional methods are available to bind these labels covalently to proteins or polypeptides. For
20 instance, coupling agents such as dialdehydes, carbodiimides, dimaleimides, bis-imidates, bis-diazotized benzidine, and the like may be used to tag the antibodies with the above-described fluorescent, chemiluminescent, and enzyme labels. See, for example, U.S. Pat. Nos. 3,940,475
25 (fluorimetry) and 3,645,090 (enzymes); Hunter et al., Nature 194:495-496, 1962; David et al., Biochemistry 13:1014-1021, 1974; Pain et al., J. Immunol. Methods 40:219-230, 1981; and Nygren, J. Histochem. and Cytochem. 30:407-412, 1982. Preferred labels are enzymes such as
30 horseradish peroxidase and alkaline phosphatase.

The conjugation of such label, including the enzymes, to the antibody is a standard manipulative procedure for one of ordinary skill in immunoassay techniques. See, for example, O'Sullivan et al., "Methods for the Preparation of
35 Enzyme-antibody Conjugates for Use in Enzyme Immunoassay," in Methods in Enzymology, ed. J.J. Langone and H. Van Vunakis, Vol. 73 (Academic Press, New York, New York, 1981), pp. 147-166. Such bonding methods are suitable for use with C-C CKR-1 or its antibodies, all of which are

proteinaceous.

Immobilization of reagents is required for certain assay methods. Immobilization entails separating the binding partner from any analyte that remains free in solution. This conventionally is accomplished by either insolubilizing the binding partner or analyte analog before the assay procedure, as by adsorption to a water-insoluble matrix or surface (Bennich et al., U.S. 3,720,760), by covalent coupling (for example, using glutaraldehyde cross-linking), or by insolubilizing the partner or analog afterward, e.g., by immunoprecipitation.

Other assay methods, known as competitive or sandwich assays, are well established and widely used in the commercial diagnostics industry.

Competitive assays rely on the ability of a tracer analog to compete with the test sample analyte for a limited number of binding sites on a common binding partner. The binding partner generally is insolubilized before or after the competition and then the tracer and analyte bound to the binding partner are separated from the unbound tracer and analyte. This separation is accomplished by decanting (where the binding partner was preinsolubilized) or by centrifuging (where the binding partner was precipitated after the competitive reaction). The amount of test sample analyte is inversely proportional to the amount of bound tracer as measured by the amount of marker substance. Dose-response curves with known amounts of analyte are prepared and compared with the test results to quantitatively determine the amount of analyte present in the test sample. These assays are called ELISA systems when enzymes are used as the detectable markers.

Another species of competitive assay, called a "homogeneous" assay, does not require a phase separation. Here, a conjugate of an enzyme with the analyte is prepared and used such that when anti-analyte binds to the analyte the presence of the anti-analyte modifies the enzyme activity. In this case, C-C CKR-1 or its immunologically active fragments are conjugated with a bifunctional organic bridge to an enzyme such as peroxidase. Conjugates are

selected for use with anti-C-C CKR-1 so that binding of the anti-C-C CKR-1 inhibits or potentiates the enzyme activity of the label. This method per se is widely practiced under the name of EMIT.

- 5 Steric conjugates are used in steric hindrance methods for homogeneous assay. These conjugates are synthesized by covalently linking a low-molecular-weight hapten to a small analyte so that antibody to hapten substantially is unable to bind the conjugate at the same time as anti-analyte.
- 10 Under this assay procedure the analyte present in the test sample will bind anti-analyte, thereby allowing anti-hapten to bind the conjugate, resulting in a change in the character of the conjugate hapten, e.g., a change in fluorescence when the hapten is a fluorophore.
- 15 Sandwich assays particularly are useful for the determination of C-C CKR-1 or C-C CKR-1 antibodies. In sequential sandwich assays an immobilized binding partner is used to adsorb test sample analyte, the test sample is removed as by washing, the bound analyte is used to adsorb
- 20 labeled binding partner, and bound material is then separated from residual tracer. The amount of bound tracer is directly proportional to test sample analyte. In "simultaneous" sandwich assays the test sample is not separated before adding the labeled binding partner. A
- 25 sequential sandwich assay using an anti-C-C CKR-1 monoclonal antibody as one antibody and a polyclonal anti-C-C CKR-1 antibody as the other is useful in testing samples for C-C CKR-1 activity.

 The foregoing are merely exemplary diagnostic assays

30 for C-C CKR-1 and antibodies. Other methods now or hereafter developed for the determination of these analytes are included within the scope hereof, including the bioassays described above.

 The following examples are offered by way of

35 illustration and not by way of limitation.

C. EXPERIMENTAL EXAMPLES

Example I

Isolation of Orphan Receptors

An "orphan receptor" cloning strategy was employed in an attempt to isolate cDNAs encoding C-C chemokine receptors. The cell surface receptors for the

5 chemoattractants C5a (Gerard, N. P. et al. Nature 349:614-7, 1991), the bacterial tripeptide fMLP (Boulay, F. et al. Biochemistry 29:11123-11133, 1990), as well as two

10 receptors for the C-X-C chemokine IL-8 receptors A and B (IL8rA and IL8rB) have been recently cloned (Holmes, W. E. et al. Science 253:1278-80, 1991; Murphy, P. M. et al. Science 253:1280-3, 1991) and found to belong to the

15 superfamily of receptor proteins whose structures are predicted to transverse the cell membrane seven times (Dohlman, H. G. et al. Annu. Rev. Biochem. 60:653-88, 1991). Since seven-transmembrane-spanning molecules are

20 typically linked to G-proteins whose function can be inhibited by pertussis toxin, we assumed that the receptors for the C-C chemokine class of proteins would also share the seven transmembrane architecture. Accordingly, two

25 degenerate oligonucleotides corresponding to conserved amino acid sequences in two transmembrane regions (TM) of the IL8rA, the C5a and the fMLP receptors were synthesized. The first oligonucleotide corresponded to a region in TM2: LNLA(L/V)AD(L/F)(L/G) (SEQ ID NO:9) and the second in TM7: NP(I/M)(I/L)Y(A/V)(F/V)(I/M/A)GQ (SEQ ID NO:10).

These oligomers were then used as primers in RT-PCR experiments using cDNA substrates from different hematopoietic cell types known to respond to C-C chemokines including peripheral blood mononuclear cells (PBMC), and

30 the cell lines U937, HL60 and THP-1. PCR was performed as follows. 1-2 µg of total RNA from different hematopoietic cell lines were used as substrates in RT-PCR (Larrick, J. W. Trends Biotech. 10:146-152, 1992), as recommended by the supplier (Perkin Elmer, Norwalk, CT). Degenerate

35 oligonucleotides corresponding to conserved regions of chemoattractant receptors were used in the PCR. PCR conditions were as follows: 94°C for 0.5', 50-55°C for 0.5' 72°C for 0.5-1', 30 cycles. PCR products were blunt-end cloned into the SmaI site of pBS (Stratagene, LaJolla, CA)

as previously described (Nguyen, T. et al. Gene 109:211-8, 1991). Plasmid DNA was isolated using the Quiagen kit (Quiagen Inc., Chatsworth, CA) as recommended by the supplier. Sequencing was performed with the Sequenase kit
5 (USBC, Cleveland, OH) as recommended by the supplier.

Subcloning and sequencing of the PCR products revealed the presence of IL8rA, IL8rB, C5a receptor and two novel clones having characteristics of seven-transmembrane-segment receptors and marked similarity to the two IL-8
10 receptors. Partial functional characterization of these orphan receptors revealed that they do not bind to HuMIP-1 α . However, it was noted that these two clones, which were more related to the IL-8 receptors than to other seven-transmembrane-spanning receptors, possessed a new
15 conserved amino acid motif at the end of TM3, DRYLAIVHA (SEQ ID NO:11), which seemed to define a subfamily of IL-8 receptor-related seven-transmembrane-spanning molecules that excluded the C5a and the fMLP receptors. Therefore, a second round of RT-PCR/orphan cloning was carried out using
20 the TM2 degenerate oligonucleotide and a DRYLAIVHA (SEQ ID NO:11) degenerate oligonucleotide. In addition, to increase the chances of obtaining C-C chemokine receptors, cDNA was obtained from cultured human monocytes, which bind radiolabeled HuMIP-1 α and from B Cells, which respond
25 chemotactically to HuMIP-1 α and used in the PCR reaction. Cloning and sequencing of these PCR products revealed several additional unique seven-transmembrane-spanning receptors.

Monocytes were cultured by standard techniques
30 commonly known in the art. In summary, buffy coat cells were separated on a Ficoll gradient. Mononuclear cells recovered from the interface were repeatedly centrifuged to remove platelets. The monocytes were separated from other mononuclear cells by adhering them to tissue culture
35 dishes. The non-adherent cells were washed off and the adherent monocytes were then were cultured for 48-72 hours before use.

Alternatively, genomic or cDNA could be screened for

the presence of orphan receptor genes by traditional Southern blot hybridization to the oligomers described above, or to probes designed from cloned DNA of a seven-transmembrane spanning protein.

5

Example II

Characterization of C-C CKR-1 DNA

A cDNA corresponding to one clone, JOSH1, was isolated by screening a λ gt10 cDNA library made from PMA (phorbol
10 12-myristate 13-acetate) treated HL60 cells with 32 P-labeled restriction fragments. Phage DNA was isolated and the inserts of the λ gt10 clones were obtained by PCR, employing 20 cycles with primers flanking the insert and the enzyme Pfu DNA polymerase (Stratagene, LaJolla, CA), as
15 recommend by the supplier. The PCR products were subcloned into the SmaI site of pRK5 as described above.

The nucleotide sequence of JOSH1 revealed an open reading frame of 1065 bases, encoding a protein of 355 amino acids (Figures 1 and 9). The deduced amino acid
20 sequence, provisionally designated as the C-C chemokine receptor 1 (C-C CKR-1), has key features related to G-protein-linked receptors of the seven-transmembrane-spanning receptor superfamily. For example, it has seven hydrophobic regions predicted to span the cell membrane,
25 and cysteine residues in the first and the second extracellular loops that are implicated in forming a disulfide bond (Figure 1). However, certain features of the predicted C-C CKR-1 protein make it distinct from classical seven-transmembrane-spanning receptors. The
30 carboxyl-terminus is relatively short and lacks cysteine residues involved in membrane anchorage via a palmitoylated moiety (O'Dowd, B. F. et al. J. Biol. Chem. 264:7564-9, 1989) and the segments between the transmembrane domains are relatively short, a feature consistent in other
35 chemoattractant receptors (Boulay, F. et al. Biochemistry 30:2993-9, 1991; Boulay, F. et al. Biochemistry 29:11123-11133, 1990; Gerard, N. P. et al. Nature 349:614-7, 1991; Holmes, W. E. et al. Science 253:1278-80, 1991; Murphy, P.

M. et al. Science 253:1280-3, 1991). There are three potential glycosylation sites in the C-C CKR-1 (Figure 1), one in the N-terminus, one in the first cytoplasmic loop and the third in TM6. This latter site is unlikely to be glycosylated since it is predicted to be embedded in the cell membrane. Finally, there is a consensus sequence for a protein kinase C phosphorylation site, at position 192, but this position is predicted to be extracellular.

The deduced amino acid sequence of C-C CKR-1 was compared to other G-protein linked chemoattractant receptors (Figure 1). The IL8rA and IL8rB showed about 32% sequence identity to C-C CKR-1, while the C5a and fMLP receptors showed about 23% identity (Figure 1). A putative seven transmembrane spanning molecule, HUMSTSR, which has been recently been deposited in Genbank (accession #M99293) by Federspiel, et al., was found to have about 31% identity with C-C CKR-1. The sequence of this molecule is identical to one of the putative receptors isolated in our first attempt at RT-PCR cloning described above. The ligand for HUMSTSR has not yet been identified. The second group of receptors which are as closely related to the C-C CKR-1 are the neuropeptide Y and the angiotensin II receptors (data not shown). Lastly, an open reading frame in the cytomegalovirus genome, designated US28, has about 33% identity with the C-C CKR-1 and about 60% identity in the N-terminal region before TM1 with C-C CKR-1.

Example III

Northern and Southern analysis of C-C CKR-1

The expression of the C-C CKR-1 was assessed in a limited panel of hematopoietic cell lines using Northern blot analysis. Northern blot hybridization was performed as follows. Total RNA was isolated using the guanidinium isothiocyanate-CsCl procedure (Sambrook et al., 1989) or by the RNazol method as recommended by the supplier. Poly A⁺ RNA was isolated using Dynabeads oligodT (DYNAL, Great Neck, NY) as recommended by the supplier. HL60 mRNA designated "HL60-C" in this report was obtained from Clontech (Palo Alto, CA). 20 µg of total RNA or 5 µg of

poly A+ RNA was fractionated on formaldehyde-agarose gels, blotted to a nitrocellulose membrane and hybridized with the C-C CKR-1 cDNA (Korneluk, R. G. et al. J. Biol. Chem. 261:8407-8413, 1986).

5 A single band of about 3 kb was detected in pre-monocytic cell lines, e.g. undifferentiated or differentiated U937 and HL60 cells (Figure 2), but not in a commercially available preparation of HL60 RNA (HL60-C, Figure 2). Lower levels of mRNA were also detected in B
10 cell lines 1788 and Daudi, but little or no RNA was detected in K562 cells (Figure 2). The highest levels were detected in PMA-treated THP-1 cells, where a strong signal was obtained when 20 mg of total RNA was analyzed (Figure 2).

15 Southern blot hybridization was performed as follows. Human genomic DNA, obtained from Clontech (Palo Alto, CA), was restriction digested, blotted onto Genescreen® (Dupont) and hybridized (Neote, K., et al. J. Clin. Invest. 86:1524-31, 1990) with the C-C CKR-1 cDNA. The hybridization
20 pattern indicated that the C-C CKR-1 gene could be intronless. Furthermore, it suggested the existence of a second related gene or possibly a pseudogene. Figures 3A and 3B represent low and high stringency washes of the same blot of human genomic DNA which has been digested with
25 several restriction enzymes. Under high stringency conditions (Figure 3B), single restriction fragments hybridizing to the C-C CKR-1 cDNA were detected when genomic DNA was digested with BamHI, HindIII or SacI and, as predicted from the cDNA sequence, two fragments are
30 detected in DNA digested with EcoRI and PstI. However, low stringency hybridization revealed the presence of additional bands that hybridized to the C-C CKR-1 cDNA e.g. an approximately 7 kb PstI fragment and an approximately 1.6 kb HindIII fragment were detected (Figure 3A). These
35 bands disappeared when the blots were washed under high stringency conditions while the other bands remained unchanged between the two washes.

Example IVSignaling through the C-C CKR-1 in response toHuMIP-1 α and RANTES

MCP-1, RANTES, HuMIP-1 α and HuMIP-1 β induce a rapid
5 and transient increase in intracellular Ca⁺⁺ in human
monocytes (Rollins, B. J. et al. Blood 78:1112-6, 1991;
Sozzani, S. et al. J. Immunol. 147:2215-21, 1991). To
determine if C-C CKR-1 was a functional C-C chemokine
receptor, it was transiently expressed in human kidney 293
10 cells and intracellular Ca⁺⁺ levels in response to
different C-C chemokines were measured.

Recombinant RANTES was expressed in E. coli and
purified as described by Kuna et al. (J. Immunol. 149:636-
42, 1992b). HuMIP-1 α and HuMIP-1 β was expressed in E.
15 coli, and purified as described by Rot et al. (J. Exp.
Med., in press). HuMIP-1 α was iodinated by the
chloroglycoluril method (Fraker et al. Biochem. Biophys.
Res. Comm. 80:849-857, 1978) to an initial specific
activity of 472 Ci/mmol. The labeled HuMIP-1 α was purified
20 by a combination of gel filtration and reversed-phase HPLC.
The ¹²⁵I-labeled RANTES, specific activity 2200 Ci/mmol,
was obtained from Dupont/NEN (Boston, MASS). Recombinant
MCP-1 and murine MIP-1 α were obtained from Peprotech (Rocky
Hill, NJ).

25 Human embryonic kidney 293 cells were transfected with
10-20 μ g of plasmid DNA by the calcium-phosphate method as
described (Schall, T. J. et al. Eur. J. Immunol. 22:1477-
81, 1992) or the modified calcium phosphate method (Chen,
C. et al. Mol. Cell. Biol. 7:2745-2752, 1987). The
30 transfected cells were assayed after transient expression
for 12-24 hours.

Intracellular Ca⁺⁺ measurements were done on the SLM
8000C essentially as described (Naccache, P. H. et al. J.
Immunol. 142:2438-44, 1989), with minor modifications:
35 IND0-1-AM, was used at 20 mg/ml final concentration and 2-4
X10⁶ cells were used per assay.

A 100 nM dose of RANTES, HuMIP-1 α , HuMIP-1 β and MCP-1 was initially used as a first approximation of the maximum physiologically-relevant concentration. When transfected cells, loaded with the calcium probe INDO-1-AM, were

5 challenged with 100 nM of either HuMIP-1 α or RANTES, a rapid increase in intracellular Ca⁺⁺ was observed (Figure 4). Challenge with the same dose of MCP-1 or HuMIP-1 β produced little detectable Ca⁺⁺ flux (data not shown). Control cells transfected with vector alone or vector

10 containing C-C CKR-1 in the opposite (non-coding) orientation did not respond to any of the ligands (data not shown). The signaling responses were dose dependent, 10 nM of HuMIP-1 α and 100 nM of RANTES were sufficient to give a maximal or near maximal response (Figure 4A and 4B).

15 Rapid, successive exposure to the same ligand is known to desensitize the signaling capacity of G-protein linked receptors (Schild, H. O. (1973), "Receptor classification with special reference to β -adrenergic receptors," In Drug Receptors, H.P. Rang, ed. (University Press), pp. 29-36).

20 In addition, desensitization can also occur when the two different agonists signal through the same receptor. HuMIP-1 α clearly blocks the ability of the C-C CKR-1 receptor to transmit a second Ca⁺⁺ signal when HuMIP-1 α is added to transfected cells twice in succession (Figure 5A).

25 Similarly, RANTES blocks the response to a second challenge at the same concentration (Figure 5B). When HuMIP-1 α is added first, it blocks the response to RANTES, indicating that complete desensitization has occurred (Figure 5C). However, challenging C-C CKR-1 cDNA transfected cells with

30 250 nM RANTES did not prevent a subsequent Ca⁺⁺ flux by the addition of 100 nM HuMIP-1 α (Figure 5D), indicating that receptor desensitization had not occurred. However, the subsequent Ca⁺⁺ flux by HuMIP-1 α is reduced, from a intracellular Ca⁺⁺ change of about 70 nM (Figure 5A and C)

35 to about 40 nM (Figure 5D), suggesting that a partial desensitization has occurred.

Example VBinding of HuMIP-1 α and RANTES to C-C CKR-1

In order to further investigate the interaction of HuMIP-1 α and RANTES to the cloned C-C CKR-1, direct binding experiments with ^{125}I -labeled ligands were carried out.

Binding assays were performed as described previously (Horuk, R. et al. J. Biol. Chem. 262:16275-16278, 1987). Transfected cells (2×10^6 cells per ml) were incubated with radiolabeled ligands and varying concentrations of unlabeled ligands at 4 °C for 2 hours. The incubation was terminated by removing aliquots from the cell suspension and separating cells from buffer by centrifugation through a silicon/paraffin oil mixture as described previously (Robb, R. J. et al. J. Exp. Med. 160:1126-1146, 1984). Non-specific binding was determined in the presence of 1 μM unlabeled ligand. Individual assay determinations, representative of at least three separate experiments were plotted. The binding data were curve fit with the computer program LIGAND (Munson, P. J. et al. Anal. Biochem. 107:220-239, 1980) modified for the IBM PC (McPherson, G. A. Comp. Prog. Biomed. 17:107-114, 1983) to determine the affinity (K_d), number of sites, and nonspecific binding. The curves shown are the binding isotherms determined by LIGAND.

Thus, when 293 cells transiently expressing C-C CKR-1 were incubated with ^{125}I -HuMIP-1 α and increasing concentrations of unlabeled HuMIP-1 α , displaceable binding of ^{125}I -HuMIP-1 α to the C-C CKR-1 was observed (Figure 6A). Scatchard analysis showed a dissociation constant (K_d) of $5.1 \pm 0.3 \text{ nM}$ and about 130000 sites/cell. This K_d is within the range of that determined for HuMIP-1 α binding to human monocytes and suggests that C-C CKR-1 is at least one of the HuMIP-1 α receptors present on monocytes. Direct binding of RANTES to the C-C CKR-1 however could not be accomplished, i.e., ^{125}I -RANTES could not be displaced by unlabeled RANTES. Interestingly, as the amount of unlabeled RANTES was increased in the binding assay, a

concomitant increase of ^{125}I -RANTES bound to cells was observed (data not shown). Since the C-C CKR-1 transduces a signal (mobilizes Ca^{++}) in response to RANTES (Figure 4), and therefore, the ligand must be binding to the cloned
5 receptor, the reason for the unusual binding profile observed for ^{125}I -RANTES is not clear. Similar binding phenomenon are obtained if other target cells responding to RANTES are used. Interestingly, ^{125}I -RANTES could be displaced by unlabeled HuMIP-1 α on 293 cells transiently
10 expressing C-C CKR-1 (Figure 6B). Scatchard analysis of this heterologous displacement suggested a K_d of $7.6 \pm 1.5 \text{ nM}$ (about 350000 sites/cell) and is consistent with the K_d of HuMIP-1 α binding data described above. These observations suggested to us initially that the C-C CKR-1 binds to
15 HuMIP-1 α and RANTES and subsequently transduces a signal by increasing the intracellular Ca^{++} levels

Example VI

Displacement of HuMIP-1 α by Heterologous Chemokines

To further characterize the binding properties of the
20 C-C CKR-1, and in particular to attempt to define a K_d for RANTES binding to the cloned C-C CKR-1, heterologous displacement of ^{125}I -HuMIP-1 α was done with RANTES and also HuMIP-1 β , MCP-1, IL-8, and murine MIP-1 α . Interestingly, all C-C chemokines (RANTES, MIP-1b and MCP-1) displaced
25 ^{125}I -HuMIP-1 α , but the C-X-C chemokine IL-8 did not (Figure 7). The K_d for RANTES, HuMIP-1b and MCP-1, and murine MIP-1 α as determined from Scatchard analysis is 468 ± 280 , 232 ± 70.1 , 122 ± 39.3 , and $4.2 \pm 2.7 \text{ nM}$ respectively, and in each case about 250000-350000 sites/cell are present
30 (Figure 7). These results revealed a broad ligand specificity of the C-C CKR-1, including some lack of species specificity in binding. More importantly, it suggested that the binding affinity between the ligand and the receptor does not predict the signaling efficacy
35 resulting from that interaction, i.e. although HuMIP-1 β and MCP-1 bind with a higher affinity than does RANTES, 100 nM of MIP-1 β or MCP-1 do not transmit a signal via the cloned receptor, whereas RANTES does (Figure 4).

AGAGAGGAAT GTAATGGTGG CCTGGGGCTT CTGAGGCTTC TGGGCTTGAG 1240
TCTTTTCCAT GAACTTCTCC CCTGGTAGAA AAGAAGATGA ATGAGCAAAA 1290
CCAAATATTC CAGAGACTGG GACTAAGTGT ACCAGAGAAG GGCTTGGACT 1340
CAAGCAAGAT TTCAGATTTG TGACCATTAG CATTTGTCAA CAAAGTCACC 1390
CACTTCCCAC TATTGCTTGC ACAAACCAAT TAAACCCAGT AGTGGTGACT 1440
GTGGGCTCCA TTCAAAGTGA GCTCCTAAGC CATGGGAGAC ACTGATGTAT 1490
GAGGA 1495

(2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 9 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

Leu Asn Leu Ala Xaa Ala Asp Xaa Xaa
1 5 9

(2) INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 10 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

Asn Pro Xaa Xaa Tyr Xaa Xaa Xaa Gly Gln
1 5 10

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 9 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

70

Asp Arg Tyr Leu Ala Ile Val His Ala
1 5 9

WE CLAIM:

1. An isolated C-C CKR-1 polypeptide.
2. A composition comprising the C-C CKR-1 of claim 1, which composition is free of a contaminating polypeptide of the animal species that is the source of C-C CKR-1 polypeptide.
3. An antibody that is capable of binding C-C CKR-1 polypeptide of claim 1 that does not cross-react with a known chemokine receptor.
4. An isolated nucleic acid molecule encoding a C-C CKR-1 polypeptide.
5. The nucleic acid molecule of claim 4 which is DNA and contains greater than about 25 bases.
6. The nucleic acid molecule of claim 4 further comprising a promoter operably linked to the nucleic acid sequence.
7. The nucleic acid molecule of claim 6, wherein the promoter is heterologous to the C-C CKR-1.
8. An expression vector comprising the nucleic acid sequence of claim 4 operably linked to control sequences recognized by a host cell transformed with the vector.

9. A host cell transformed with the vector of claim 8.
10. A method of determining the presence of a C-C CKR-1 nucleic acid, comprising hybridizing nucleic acid encoding or complementary to C-C CKR-1 nucleic acid to a test sample nucleic acid and determining the presence of C-C CKR-1 nucleic acid.
11. A method of amplifying a nucleic acid test sample comprising priming a nucleic acid polymerase reaction with nucleic acid encoding or complementary to a C-C CKR-1 nucleic acid.
12. A composition comprising the C-C CKR-1 polypeptide of claim 1 and a pharmaceutically acceptable carrier.
13. A composition comprising the antibody of claim 3 and a pharmaceutically acceptable carrier.
14. An isolated polypeptide comprising the C-C CKR-1 polypeptide of claim 1 fused to a polypeptide heterologous to the C-C CKR-1.
15. A DNA isolate able to hybridize under stringent conditions to a nucleotide sequence encoding C-C CKR-1 having the amino acid sequence shown in Figure 1 and encoding a polypeptide having C-C CKR-1 biological activity.
16. The DNA isolate of claim 15 wherein the DNA isolate